

## Research Article

# Additions to the genus *Mycena* (Mycenaceae, Agaricales): Descriptions of five new taxa in Hunan Province, China

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#### **Abstract**

Few studies have been conducted on *Mycena* species in Hunan Province, China. During our research on the species diversity of *Mycena* in Hunan Province, we identified approximately 30 *Mycena* species based on morphological and molecular evidence. Five species are recognized herein as new to science, namely, *M. fulvomarginata*, *M. huang-sangensis*, *M. hongfengensis*, *M. subroriduliformis*, and *M. roseolamellata*. The phylogenetic analyses of a combined ITS and LSU sequence dataset revealed that five new species each formed an independent lineage that could separate phenotypically similar and phylogenetically related species. Descriptions, photographs, and phylogenetic analysis results are provided for the five new species, along with the comparisons with related species. A key to all *Mycena* species found in Hunan is also provided.

Key words: Basidiomycota, biodiversity, five new species, phylogenetic analysis, taxonomy

# Introduction

Hunan Province is located in the southern middle reaches of the Yangtze River, China, and covers an area of approximately 211,800 km² (Gao and Dou 1981). The province is surrounded by mountains and hills in the east, west, and south, basins in the center, and plains in the north. This area has a subtropical humid monsoon climate with four distinct seasons, sufficient sunshine, and abundant rainfall, which benefits vegetation (Qing 1990). The people's government of Hunan Province reported that there are 17 national nature reserves, including 16 focused on preserving forest ecological systems (https://www.hunan.gov.cn/). The unique topography and good forest ecological systems make Hunan home to macrofungi (Liu et al. 2024), among which the members of *Mycena* (Pers.) Roussel are prominent in Agaricales (Liu et al. 2022a, 2022b).

Mycena is one of the largest genera in the Mycenaceae family of Agaricales, including at least 600 species worldwide (Kirk et al. 2008; Fan et al. 2024; Zhang

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et al. 2024). *Mycena* plays an important role in ecosystems, which depend on the strong decomposition abilities of fungi to promote material circulation, forest metabolism, and natural renewal (Fukasawa et al. 2009; Guerreiro et al. 2023). Some *Mycena* are symbiotic fungi with *Gastrodia elata* seeds (Lee et al. 2017; Liu et al. 2022b, 2025). Moreover, approximately 60 *Mycena* species are reported to be bioluminescent worldwide (Chew et al. 2014, 2015; Cortés-Pérez et al. 2019, 2023; Lu et al. 2024).

Mycena was first moved to the genus level by Roussel in 1806, but this change was not widely accepted by the majority of mycologists until the 20<sup>th</sup> century (Roussel 1806). It is a widely studied genus despite many species in the genus having small basidiomata (Aronsen and Læssøe 2016). Many important contributions about taxonomic studies of Mycena have been made to temperate regions, and several monographs have been published (Smith 1947; Robich 2003, 2016; Aronsen and Læssøe 2016). For subtropical and tropical areas, the study of local floras has contributed to the early reporting of Mycena species (Métrod 1949; Singer and Digilio 1951; Pegler 1977, 1983, 1986, 1987), and those studies made substantial contributions to more important papers and related monographs on Mycena (Singer 1989; Corner 1994; Grgurinovic 1998, 2003; Perry 2002; Cooper et al. 2018; Bau et al. 2021).

Some classifications have been proposed based on the morphological characteristics of Mycena (Fries 1821; Lange 1914; Kühner 1926, 1931, 1938; Singer and Digilio 1951; Singer 1986; Maas Geesteranus 1992a, 1992b; Maas Geesteranus and Horak 1995; Bau et al. 2021). Currently, mycologists tend to accept, use, and update the infrageneric classification, which was proposed by Maas Geesteranus (Grgurinovic 1998, 2003; Robich 2003, 2016; Cooper et al. 2018). Species are classified into 44 sections based on a combination of macroscopic and microscopic features. With the development of molecular biology research, the phylogenetic positions of *Mycena* have become increasingly apparent (Moncalvo et al. 2002; Matheny et al. 2006; Wei et al. 2024). As a part of the subtropical area of China, studies on Mycena have been conducted in Hunan Province. The book Hunan Macrofungi was specially written in 1997 to document the fungi in the region and described two Mycena species (Li et al. 1993). Five Mycena species from Hupingshan, Hunan, are described in the Atlas of Macrofungi (Zhang et al. 2005). Seven Mycena species were recorded in the Atlas of Macrofungi in Hunan (Chen and Zhang 2019). A species in China, M. heteracantha (Singer) Desjardin, which was collected from Hunan Province, was described by Na Q and Bau T in 2019 (Na and Bau 2019a). Mycena subpiligera L.N. Liu, a new species collected in Hunan Province that can significantly enhance the germination efficiency of Gastrodia elata seeds, was reported in 2022 (Liu et al. 2022b). Mycena chlorocyanea L.N. Liu, another new species whose type was collected in Hunan Province, was reported in recent years (Liu et al. 2022a). As a region that is rich in natural resources, further studies on *Mycena* in Hunan Province are needed.

Aiming to explore the diversity of *Mycena* in the region, we conducted literature reviews and field investigations in Hunan Province from 2020 to 2024. Through macroscopic comparison and phylogenetic analysis, we found 30 *Mycena* species collected from Hunan Province, including five new species that are described in this paper.

## Materials and methods

# Sample collection and morphological description

Approximately 400 Mycena specimens belonging to 30 taxa were collected from Hunan Province (Table 1). During fieldwork, the collected samples were photographed, and additional information, such as elevation, habitat, and collection date, was recorded (Rathnayaka et al. 2024). Detailed morphological features, including basidiomata size, color and shape, odor, taste, and viscosity, were also documented from fresh specimens. The specimens were dried with silica gel and deposited in the Herbarium of Hunan Institute of Forestry (HUIF) and the Herbarium of Jishou University (JSU). The macromorphological characteristics of the samples were determined based on field notes and photographs. The color codes followed Kornerup & Wanscher (Kornerup and Wanscher 1978). Micromorphological characters of dry specimens were observed under light microscopy (Olympus BX51). To measure the sizes of related tissues, basidiospores, basidia, pileipellis, stipitipellis, and other tissues were mounted in pure water, 5% KOH solution, or 1% Congo Red solution. Melzer's reagent was used to test the amyloid reaction of the spores. At least 30 basidiospores were measured in each sample. The Q value refers to the length/ width ratio of the basidiospores. Q ± av represents the average Q of all basidiospores ± the sample standard deviation. The ranges of the basidiospores are presented as (a-) b-c (-d): the range 'b-c' represents 90% of the measured values, while 'a' and 'd' represent extreme values. The measurements of other microscopic structures were based on 20 measurements per specimen.

# DNA extraction, PCR amplification, and sequencing

Genomic DNA was extracted from fresh or dried specimens using the NuClean Plant Genomic DNA kit (Kangwei Century Biotechnology Co., Beijing, China) following the manufacturer's protocols. The ITS rDNA region (ITS1–5.8S–ITS2) was amplified using the primer pair ITS1 and ITS4 (White et al. 1990). The LSU region was amplified with the primers LR0R and LR7 (Vilgalys and Hester 1990). PCR was performed in a total volume of 25  $\mu$ L containing 1  $\mu$ L template DNA, 9.5  $\mu$ L distilled water, 1  $\mu$ L of each primer, and 12.5  $\mu$ L 2x Taq PCR Master Mix with blue dye (Sangon Biotech, Shanghai, China). The PCR conditions were as follows: initial denaturation at 95 °C for 5 min, followed by 35 cycles of 94 °C for 45 s, 45 s at 52 °C, and 1 min at 72 °C for ITS (Liu et al. 2022a). For the LSU conditions: initial denaturation at 95 °C for 4 min, followed by 35 cycles of 95 °C for 1 min, 1 min at 53 °C, and 80 s at 72 °C (Zhang et al. 2024). The amplified products were determined by electrophoresis on a 1% agarose gel against a known standard DNA marker and directly sequenced at Sangon Biotech. Newly generated sequences in this study have been submitted to the NCBI GenBank database.

# Molecular phylogeny

Details of the sequences used for phylogenetic analysis were obtained from this study and downloaded from GenBank (Table 2). DNA sequences were checked using Bioedit v7.0.9 to ensure sequencing quality (Hall 1999). SeqMan 7.1.0 was used for splicing and manual editing (Swindell and Plasterer 1997).

**Table 1.** Summary of the collection of *Mycena* species and their distributions and seasons in Hunan.

Species name	Ecology and location (in Hunan)	Occurrence time (in Hunan)	Geographical compositions	References
M. abramsii	Solitary, in small groups or subfasciculate on dead twigs or woody debris of deciduous trees; also occurs occasionally on dead twigs of coniferous trees. Widespread in Hunan.	May to July	Worldwide distribution	(Singer and Digilio 1951; Hintikka 1963; Watling 1984; Maas Geesteranus 1988; Emmett 1992; Marra 2000; Tholl et al. 2000; Veerkamp 2001; Laganà et al. 2002; Perry 2002; Robich 2003; Afyon et al. 2005; Bronckers and Kelderman 2011; Senn-Irlet et al. 2012; Gierczyk et al. 2015; Aronsen and Læssøe 2016; Rudolf et al. 2016; Vishwakarma et al. 2017; Bau et al. 2021)
M. adnexa	In small groups on dead twigs of deciduous trees. Widespread in Hunan.	May to June	Worldwide distribution	(Bau et al. 2021)
M. algeriensis	Subfasciculate on rotten stumps in deciduous forests. Collected only in western areas of Hunan.	April to May	Temperate-subtropical and tropical distribution	(Hintikka 1963; Laganà et al. 2002; Gierczyk et al. 2015; Aronsen and Læssøe 2016; Bau et al. 2021)
M. bicystidiata	Scattered on rotten wood in mixed forests. Collected in western and eastern areas of Hunan.	April to June	Only found in China	(Bau et al. 2021)
M. castaneicola	Scattered or gregarious on Castanea mollissima fruits. Collected only in western areas of Hunan.	June to July	Only found in China	(Bau et al. 2021)
M. chlorocyanea	Gregarious in leaf humus under deciduous trees. Collected only in western areas of Hunan.	April to May	Only found in China	(Liu et al. 2022a)
M. citrinomarginata	Gregarious in leaf humus under deciduous trees. Collected in western and eastern areas of Hunan.	May to June	Temperate-subtropical and tropical distribution	(Smith 1935; Hintikka 1963; Watling 1984; Senn-Irlet 1987; Emmett 1992; Laganà et al. 2002; Perry 2002; Kalamees and Raitviir 2006; Senn-Irlet et al. 2012; Gierczyk et al. 2015; Gáperová et al. 2015; Na and Bau 2018; Lee et al. 2021)
M. corynephora	Gregarious on bark of living deciduous trees. Widely distributed in Hunan.	April to May	Worldwide distribution	(Desjardin 1995; Laganà et al. 2002; Aronsen and Læssøe 2016; Bau et al. 2021)
M. deeptha	Gregarious on rotten wood of living deciduous trees. Collected in western and southern areas of Hunan.	April to July	Temperate-subtropical and tropical distribution	(Aravindakshan et al. 2012)
M. digitifurcata	Gregarious on bark of living deciduous trees. Collected only in Changsha.	June	Only found in China	(Wei et al. 2024)
M. filopes	Solitary on dead twigs of deciduous trees. Collected only in western areas of Hunan.	October	Worldwide distribution	(Beardslee and Coker 1924; Watling 1984; Kalamees and Raitviir 2006; Bronckers and Kelderman 2011; Senn-Irlet et al. 2012; Aronsen and Læssøe 2016; Bau et al. 2021; Cho and Chung 2023)
M. fulvomarginata	Gregarious on moss-covered bark of living deciduous trees. Collected only in western areas of Hunan.	June to July	Only found in China	This study
M. galericulata	Solitary to fasciculate on branches, trunks and stumps of deciduous trees. Collected only in western areas of Hunan.	October to November	Worldwide distribution	(Beardslee and Coker 1924; Swartz 1933; Imai 1938; Hintikka 1963; Ballesteros 1984; Watling 1984; Maas Geesteranus 1992c; Nauta and Vellinga 1992; Marra 2000; Tholl et al. 2000; Laganà et al. 2002; Perry 2002; Robich 2003; Senn-Irlet et al. 2012; Gáperová et al. 2015; Aronsen and Læssøe 2016; Gyosheva et al. 2016; Vishwakarma et al. 2017; Cho and Chung 2020, 2023; Bau et al. 2021; Lee et al. 2021; Luszczyński et al. 2022)
M. haematopus	In small groups or fasciculate on twigs and trunks of deciduous trees. Distributed in eastern, western and southern areas of Hunan.	May, June, October, November	Worldwide distribution	(Beardslee and Coker 1924; Swartz 1933; Imai 1938; Hintikka 1963; Watling 1984; Emmett 1992; Grgurinovic 1998; Laganà et al. 2002; Perry 2002; Robich 2003; Bronckers and Kelderman 2011; Senn-Irlet et al. 2012; Aravindakshan and Manimohan 2013b; Aronsen and Læssøe 2016; Mustafabayli and Aghayeva 2019; Bau et al. 2021; Cho and Chung 2023)

Species name	Ecology and location (in Hunan)	Occurrence time (in Hunan)	Geographical compositions	References
M. heteracantha	Gregarious on decaying leaves and twigs of deciduous trees. Collected only in southwestern areas of Hunan.	May	Temperate-subtropical and tropical distribution	(Desjardin 1995; Bau et al. 2021)
M. hongfengensis	Gregarious on decaying leaves of deciduous trees. Collected only in western areas of Hunan.	April	Only found in China	This study
M. huangsangensis	Gregarious on decaying leaves of deciduous trees. Collected only in southwestern areas of Hunan.	April to May	Only found in China	This study
M. laevigata	In small groups or fasciculate on twigs and trunks of deciduous trees. Collected in western and southern areas of Hunan.	June to September	Temperate-subtropical and tropical distribution	(Chen and Zhang 2019)
M. leaiana	Fasciculate on rotten wood of deciduous trees. Collected in western and southern areas of Hunan.	July	Temperate-subtropical and tropical distribution	(Chen and Zhang 2019)
M. leptocephala	Gregarious on moss-covered hardwood or on branches. Collected in western and southern areas of Hunan.	April to May	Worldwide distribution	(Beardslee and Coker 1924; Smith 1935; Hintikka 1963; Watling 1984; Senn-Irlet 1987; Laganà et al. 2002; Kalamees and Raitviir 2006; Bronckers and Kelderman 2011; Senn-Irlet et al. 2012; Gierczyk et al. 2015; Aronsen and Læssøe 2016; Baldrian et al. 2016; Bau et al. 2021; Łuszczyński et al. 2022)
M. maculata	Solitary to fasciculate on branches, trunks and stumps of deciduous trees. Collected in western areas of Hunan.	October to November	Temperate distribution	(Marra 2000; Tholl et al. 2000; Laganà et al. 2002; Perry 2002; Bau et al. 2021)
M. meliigena/ juniperina	Gregarious on moss-covered bark of living deciduous trees. Widespread.	November to June	Temperate distribution	(Aronsen 1996; Doğan and Karadelev 2006; Halama et al. 2014)
M. pearsoniana	Scattered in leaf humus in deciduous trees. Collected only in western areas of Hunan.	May to June	Worldwide distribution	(Hintikka 1963; Watling 1984; Moreno and Albertó 1996; Robich 2003; Senn-Irlet et al. 2012; Aronsen and Læssøe 2016; Türkekul 2017; Vishwakarma et al. 2017; Kwon et al. 2020; Bau et al. 2021)
M. picta	Scattered on decaying leaves of deciduous trees. Collected only in some parks of Changsha.	April to May	Temperate-subtropical and tropical distribution	(Miyamoto et al. 1996; Halama and Romański 2010; Shiryaeva 2018; Retnowati et al. 2020; Bau et al. 2021)
M. pluteoides	Solitary or gregarious on rotten wood of deciduous trees. Collected in western and southern areas of Hunan.	May, June	Only found in China	(Bau et al. 2021)
M. pura	Scattered in leaf humus and on needles or in grasslands, on both deciduous and coniferous trees. Widely distributed in Hunan.	November, March to June	Worldwide distribution	(Beardslee and Coker 1924; Swartz 1933; Imai 1938; Hintikka 1963; Watling 1984; Senn-Irlet 1987; Emmett 1992; Maas Geesteranus 1992c; Marra 2000; Laganà et al. 2002; Perry 2002; Robich 2003; Kalamees and Raitviir 2006; Senn-Irlet et al. 2012; Casabón 2015; Gáperová et al. 2015; Aronsen and Læssøe 2016; Gyosheva et al. 2016; Mustafabayli and Aghayeva 2019; Bau et al. 2021; Lee et al. 2021)
M. roseolamellata	Gregarious on decayed twigs of bamboo or woody debris of deciduous trees. Ningxiang, Hunan.	November, December and March	Only found in China	This study
M. subpiligera	Longshan and Suining Counties, Hunan.	April to July	Only found in China	(Liu et al. 2022b)
M. subroriduliformis	Gregarious on decaying leaves of deciduous trees. Suining County, Hunan.	April to May	Only found in China	This study
M. yuezhuoi	Scattered on the litter layer in Pinus, Quercus, and Robinia mixed forests. Suining County, Hunan.	April to May	Temperate-subtropical and tropical distribution	(Liu et al. 2021; Cho et al. 2024)

**Table 2.** Names, voucher numbers, locations, and corresponding GenBank accession numbers of the taxa used in the phylogenetic analysis. - refers to the data unavailability.

Species	Voucher	GenBank ad	ccession no.	Location	References	
Species	Voucher	ITS	LSU	Location		
Atheniella adonis	H6036863	MW540691	-	Finland	Unpublished	
A. aurantiidisca	UBCF33062	MF908459	-	Canada	Unpublished	
Clitocybula intervenosa	BAP 588	MH414560	-	Africa	(Cooper et al. 2018)	
C. intervenosa	BAP 613	MH414561	MH385335	Africa	(Cooper et al. 2018)	
lydropus murinus	BAP 657	MH414565	-	Africa	(Cooper et al. 2018)	
Mycena abramsii	HUIFS50116	OP604436	OP605596	China	Unpublished	
M. abramsii	HUIFS50074	OP604427	-	China	Unpublished	
М. abramsii	HUIF50533	PQ406957	-	China	This study	
M. adnexa	HMAJU43360	MK733290	MK722345	China	Unpublished	
M. adnexa	HMAJU43691	MK733293	MK722346	China	Unpublished	
M. adnexa	HUIF50339	PQ406958	-	China	This study	
M. adnexa	HUIF60005	PQ465300	-	China	This study	
M. albiceps	F27622	MZ303026	-	USA	Unpublished	
M. albiceps	RA705-6	MK234177	-	USA	Unpublished	
M. algeriensis	HMAS 291753	OR236986	-	China	Unpublished	
M. algeriensis	HUIF50368	PQ406959	-	China	This study	
M. alniphila	904	JF908482	-	Italy	Unpublished	
M. amicta	CBS:254.53	MH857183	_	France	(Vu et al. 2019)	
M. amicta M. amicta	CBS:352.50	MH856655	MH868170	France	(Vu et al. 2019)	
M. arcangeliana	252b	JF908401	10111000170	Spain	(Osmundson et al. 2013)	
	252f	-	-		<u> </u>	
M. arcangeliana		JF908402	- M//6000E0	Spain	(Osmundson et al. 2013	
M. bicystidiata	HMJAU43648	MK309773	MK629359	China	(Na and Bau 2019b)	
M. bicystidiata	HUIF50044	PQ406952	-	China	This study	
M. bicystidiata	HUIF50583	PQ406953	-	China	This study	
M. breviseta	BAP 633	MH414551	MH385327	Africa	(Cooper et al. 2018)	
M. brunnescens	JSU125	ON778578	OP360941	China	(Zhang et al. 2024)	
M. brunnescens	JSU126	ON778579	OP360942	China	(Zhang et al. 2024)	
M. brunnescens	JSU127	PP152232	-	China	(Zhang et al. 2024)	
M. bulliformis	SFSU:BAP 547	KX513844	KX513848	USA	(Perry and Desjardin 201	
M. caeruleomarginata	FFAAS0358	OL711670	OL711665	China	(Na et al. 2022)	
M. caeruleomarginata	FFAAS0357	OL711669	OL711664	China	(Na et al. 2022)	
M. capillaripes	HRL2854	PQ811198	-	USA	Unpublished	
M. castaneicola	JSU138	PQ406949	-	China	This study	
M. castaneicola	JSU263	PQ406950	-	China	This study	
M. castaneicola	HMJAU43581	MH136827	-	China	(Na and Bau 2019a)	
M. cf. cinerella	173	MF926553	-	Russia	(Malysheva et al. 2017)	
M. chlorocyanea	HUIF50234	OP358280	OP360937	China	(Liu et al. 2022a)	
M. chlorocyanea	HUIF50238	OP358281	OP360938	China	(Liu et al. 2022a)	
M. chlorophos	CT15101401	MH400938	-	China	Unpublished	
M. cinerella	Aronsen051014	KT900146	-	Norway	Unpublished	
M. citrinomarginata	SHXG	OM228755	OM228763	China	Unpublished	
M. citrinomarginata	HMJAU 43563	MG654739	-	China	(Na and Bau 2018)	
M. confinationis	MO362993	PP831662	-	USA	Unpublished	
M. confinationis	PAMP-fungi-41	MT764847	MT764850	Spain	Unpublished	
Л. corynephora	JSU145	PQ406951	-	China	This study	
 И. corynephora	SJiao	OP604434	-	China	Unpublished	
M. cristinae	JS347	MT921381	MT921384	Brazil	(Oliveira et al. 2021)	
M. cristinae	JS767	MT921382	-	Brazil	(Oliveira et al. 2021)	
M. crocea	S.D. Russell iNaturalist #16588497	OM473679	-	USA	Unpublished	
M. crocea	OMDL K. Canan iNaturalist 182892200	PP436589	-	USA	Unpublished	
M. cyanorhiza	J24082010	MW540696	-	Finland	Unpublished	
M. cyanorhiza M. cyanorhiza	120b	JF908385	-	Italy	(Osmundson et al. 2013	

Species	Voucher	GenBank ad	cession no.	Location	References	
Species	Voucilei	ITS	LSU	Location	References	
M. deeptha	DM334g (K(M)178333)	JX481737	-	India	(Aravindakshan et al. 2012)	
M. deeptha	HUIF50518	PQ406962	-	China	This study	
M. digitifurcata	HUIF60006	PQ406940	-	China	(Wei et al. 2024)	
M. digitifurcata	FFAAS1055	PP706100	PP704700	China	(Wei et al. 2024)	
M. entolomoides	HMJAU 43126	MG654738	-	China	(Na and Bau 2018)	
M. entolomoides	HMJAU 43052	MG654737	-	China	(Na and Bau 2018)	
M. entolomoides	HMJAU 43048	MG654736	-	China	(Na and Bau 2018)	
M. filopes	HUIF50198	OP604441	OP605599	China	Unpublished	
M. filopes	HMAS 291835	OR236988	-	China	Unpublished	
M. flosoides	HUIF50128	OP358282	OP360939	China	(Liu et al. 2022a)	
M. flosoides	HUIF50129	OP358283	OP360940	China	(Liu et al. 2022a)	
M. flosoides	HUIF50128-R	OP745013	-	China	(Liu et al. 2022a)	
M. fulgoris	ACP1690	MG926694	-	Mexico	(Cortés-Pérez et al. 2019)	
M. fulgoris	ACP1785	MG926693	-	Mexico	(Cortés-Pérez et al. 2019)	
M. fulvomarginata	HUIF50088 Holotype	PQ406943	-	China	This study	
M. fulvomarginata	HUIF50089	PQ406944	PQ406964	China	This study	
M. galericulata	TFB14675	MN088380	-	USA	(Hughes et al. 2020)	
M. galericulata	TFB14649	MN088382	-	USA	(Hughes et al. 2020)	
M. galericulata	HUIF50196	OP604439	-	China	Unpublished	
M. haematopus	HUIF50203	OP604443	OP605601	China	Unpublished	
M. haematopus	HMJAU43662	MK733299	MK722353	China	Unpublished	
M. huangsangensis	HUIF50526 Holotype	PQ406935	-	China	This study	
M. huangsangensis	HUIF50528	PQ406936	PQ406965	China	This study	
M. interrupta	HMJAU43849	MK733301	-	China	Unpublished	
M. interrupta	HMJAU43791	MK733300	-	China	Unpublished	
M. juniperina	869	JF908478	-	Italy	(Osmundson et al. 2013)	
M. laevigata	HMJAU43618	MK733304	MK722355	China	Unpublished	
M. laevigata	MHHNU 8626	MK453048	-	China	Unpublished	
M. leaiana	MHHNU 30544	MK250916	<u> </u>	China	Unpublished	
M. leaiana	HKAS126400	OQ025147		China	Unpublished	
M. leptocephala	HUIF50005	PQ406956	_	China	This study	
M. leptocephala	CA FUNDIS iNaturalist #160824125	OR778420	_	USA	Unpublished	
M. longinqua	BAP 648	MH414552	MH385328	Africa	(Cooper et al. 2018)	
M. maculata	HUIFS50209	OP604446	-	China	Unpublished	
M. maculata	HMJAU43009	MK309791	MK629347	China	Unpublished	
M. meliigena	39c	JF908428	WIN029347		(Osmundson et al. 2013)	
	390	JF908423		Italy		
M. meliigena			-	Italy	(Osmundson et al. 2013)	
M. meliigena/ juniperina	HUIF60003	PQ406954	-	China	This study	
M. meliigena/ juniperina	HUIF60004	PQ406955	-	China	This study	
M. metata	313b	JF908412	-	Italy	(Osmundson et al. 2013)	
M. metata	HMJAU43625	MH396636	-	China	Unpublished	
M. hongfengensis	JSU114 Holotype	PQ406945	PQ406967	China	This study	
M. hongfengensis	JSU121	PQ406946	PQ406968	China	This study	
M. oryzifluens	FFAAS1051	PP706096	PP704696	China	(Wei et al. 2024)	
M. pasvikensis	AAronsen50-13	KU861558	-	Norway	Unpublished	
M. pasvikensis	AAronsen86-12	KU861556	-	Norway	Unpublished	
M. pearsoniana	TENN61384	JN182200	-	USA	(Harder et al. 2012)	
M. pearsoniana	TENN61544	JN182199	-	USA	(Harder et al. 2012)	
M. pearsoniana	HUIF50392	PQ406948	-	China	This study	
M. picta	CA FUNDIS iNaturalist 171114596	OR858681	-	USA	Unpublished	
M. picta	TUR194167	MW540717	-	Finland	Unpublished	
M. pluteoides	HMJAU43771	MK733307	MK722357	China	Unpublished	
M. pluteoides	HMJAU43765	MK733306	-	China	Unpublished	
M plutopidos	HUIF50584	PQ406961	_	China	This study	
M. pluteoides	11011 30304	1 0 100501			· · · · · · · · · · · · · · · · · · ·	

Species	Voucher	GenBank accession no.		Location	References
Species	voucher	ITS	LSU	Location	References
M. polygramma	CBS:240.47	MH856235	MH867764	France	(Vu et al. 2019)
M. polygramma	439b	JF908433	-	Italy	(Osmundson et al. 2013)
M. pura	HUIF50006	OP604419	OP605597	China	Unpublished
M. pura	TENN60139	EU517505	-	Russia	(Petersen et al. 2008)
M. purpureofusca	HMJAU 43554	MG654740	-	China	(Na and Bau 2018)
M. purpureofusca	HMJAU 43624	MG654741	-	China	(Na and Bau 2018)
M. rosella	73h	JF908471	-	Italy	(Osmundson et al. 2013
M. rosella	53	MW576937	-	Norway	Unpublished
M. roseolamellata	HUIF60001 Holotype	PQ406941	PQ406969	China	This study
M. roseolamellata	HUIF60002	PQ406942	-	China	This study
M. rubromarginata	CBS:265.48	MH856335	MH867890	France	(Vu et al. 2019)
M. rubromarginata	CBS:268.48	MH856338	MH867891	France	(Vu et al. 2019)
M. sanguinolenta	TENN59879	FJ596764	-	USA	(Hughes et al. 2009)
M. seynesii	71h	JF908470	-	Italy	(Osmundson et al. 2013)
M. seynesii	711	JF908469	-	Italy	(Osmundson et al. 2013
Mycena sp.	JSU008	PQ465299	-	China	Unpublished
Mycena sp.	JSU132	PQ406963	-	China	This stufy
Mycena sp.	080108	LC504829	-	Japan	This study
M. silvaenigrae	HMJAU43815	MK733310	MK722359	China	Unpublished
M. subcaerulea	TENN-F-051121	OL711671	OL711666	USA	(Na et al. 2022)
M. subcaerulea	TENN-F-057919	OL711672	OL711667	USA	(Na et al. 2022)
M. subpiligera	HUIF50036	OM228758	-	China	(Liu et al. 2022b)
M. subpiligera	HUIFS50007	OM228759	-	China	(Liu et al. 2022b)
M. subroriduliformis	HUIF50540 Holotype	PQ406937	PQ406970	China	This study
M. subroriduliformis	HUIF50546	PQ406938	<u>-</u>	China	This study
M. substylobates	HMJAU43444	MH216190	-	China	(Na and Bau 2019a)
M. substylobates	HMJAU43418	MH216189	-	China	(Na and Bau 2019a)
M. tenax	OSC 113746	EU846251	-	USA	Unpublished
M. tenax	OSC 113728	EU669224	-	USA	Unpublished
M. vulgaris	447h	JF908435	-	Italy	Unpublished
M. vulgaris	CBS:248.47	MH856240	MH867770	France	(Vu et al. 2019)
M. xantholeuca	CBS370.50	MH856663	MH868180	France	(Vu et al. 2019)
M. xantholeuca	CBS371.50	MH856664	MH868181	France	(Vu et al. 2019)
M. yuezhuoi	FFAAS0346	MW581492	-	China	(Liu et al. 2021a)
M. yuezhuoi	HUIF50535	PQ406947	-	China	(Liu et al. 2021)
M. zephirus	CBS:270.48	MH856339	MH867892	France	(Liu et al. 2021)
M. zephirus	AH60146	PP868143	-	Spain	(Villarreal et al. 2024)
Phloeomana minutula	H6036841	MW540684	-	Finland	Unpublished
P. speirea	iNAT: 100003394	ON206666	-	USA	Unpublished

The final datasets were aligned using MAFFT v.7.310 (Katoh and Standley 2013). The sequences were concatenated into one multi-loci dataset with SequenceMatrix 1.7.8 (Vaidya et al. 2011). The ALTER (Alignment Transformation EnviRonment) online tool was used for the final conversion of the FASTA format to the NEXUS format (Glez-Penñ et al. 2010). The best-fit evolutionary model was selected using MrModelTest v.2.3 under the Akaike information criterion (AIC) (Nylander 2004). A phylogenetic tree was constructed based on maximum likelihood and Bayesian inference methods. Maximum likelihood (ML) analyses were performed with RAxML-NG v.0.9.0 (Kozlov et al. 2019), and bootstrap values were calculated from 1,000 replicates. Bayesian inference analysis was performed using the Metropolis-coupled Markov chain Monte Carlo method with MrBayes v3.2.5 under the GTR +I+G model (Ronquist and Huelsenbeck 2003). Analyses were run with 4 chains of 2,000,000 generations, and trees

were sampled every 100<sup>th</sup> generation. The first 25% of the sample trees were discarded as burn-in. Gaps were treated as missing data. Phylogenetic trees were visualized with FigTree v1.4.3 (http://tree.bio.ed.ac.uk/software/figtree/).

# **Results**

# Phylogenetic analysis

The two-locus dataset (ITS + LSU) consisted of 191 sequences and 1,680 nucleotide sites in total, which are shown in Table 2. It includes sequences of 28 *Mycena* taxa except for *M. picta* (Fr.) Harmaja and *M. heteracantha*, which are present in Hunan. Sequences of closely related species with high homology and morphologically similar species were also downloaded from GenBank. *Atheniella adonis* (Bull.) Redhead, Moncalvo, Vilgalys, Desjardin, & B.A. Perry, *A. aurantiidisca* (Murrill) Redhead, Moncalvo, Vilgalys, Desjardin, & B.A. Perry; *Clitocybula intervenosa* A.C. Cooper, Desjardin, & B.A. Perry (BAP 588, BAP 613), *Phloeomana minutula* (Sacc.) Redhead; *P. speirea* (Fr.) Redhead, and *Hydropus murinus* A.C. Cooper, Desjardin, & B.A. Perry were chosen as the outgroup (Liu et al. 2022a). The topologies generated from maximum likelihood (ML) and Bayesian inference (BI) analyses were identical, although statistical support for some branches showed slight differences. The BI tree with branch lengths inferred from the ITS and LSU datasets is shown in Fig. 1.

The phylogeny inferred from the combined dataset revealed that the *Mycena* split into two well-supported clades, and all new taxa formed a well-supported monophyletic lineage. *Mycena hongfengensis* formed a small branch and grouped with an unidentified *Mycena* sp. in clade 1 (BS/BP = 100/1.00). *Mycena roseolamellata*, *M. fulvomarginata*, *M. huangsangensis*, and *M. subroriduliformis* were members of clade 2. *Mycena roseolamellata* and *M. entolomoides* T. Bau formed a supported branch in the tree (BS/BP = 100/1.00), and their genetic distance is substantial enough to distinguish between the two species. *Mycena fulvomarginata* is most related to *M. capillaripes* Peck. They grouped together with BS/BP = 83/1.00 statistical support, and they were distinct. *Mycena albiceps* (Peck) Gilliam, *M. flosoides* L.N. Liu, *M. brunnescens* L.N. Liu, and our specimens (*M. huangsangensis* and *M. subroriduliformis*) formed a separate branch with strong statistical support (BS/BP = 86/1.00).

#### **Taxonomy**

Mycena huangsangensis L.N. Liu, sp. nov.

MycoBank No: 856016

Figs 2, 3

**Diagnosis.** Differs from the most similar species, *M. alniphila*, by its decurrent lamellae and longer basidiospores.

**Holotype.** CHINA • Hunan Province, Shaoyang City, Suining County, Hunan Huangsang National Nature Reserve, 26°24'18"N, 110°05'37"E, elev. 644 m, 24 April 2024, LiNa Liu, *HUIF50526* (collection number NN526).

**Etymology.** Refers to the Huangsang National Nature Reserve, from where the holotype was collected.

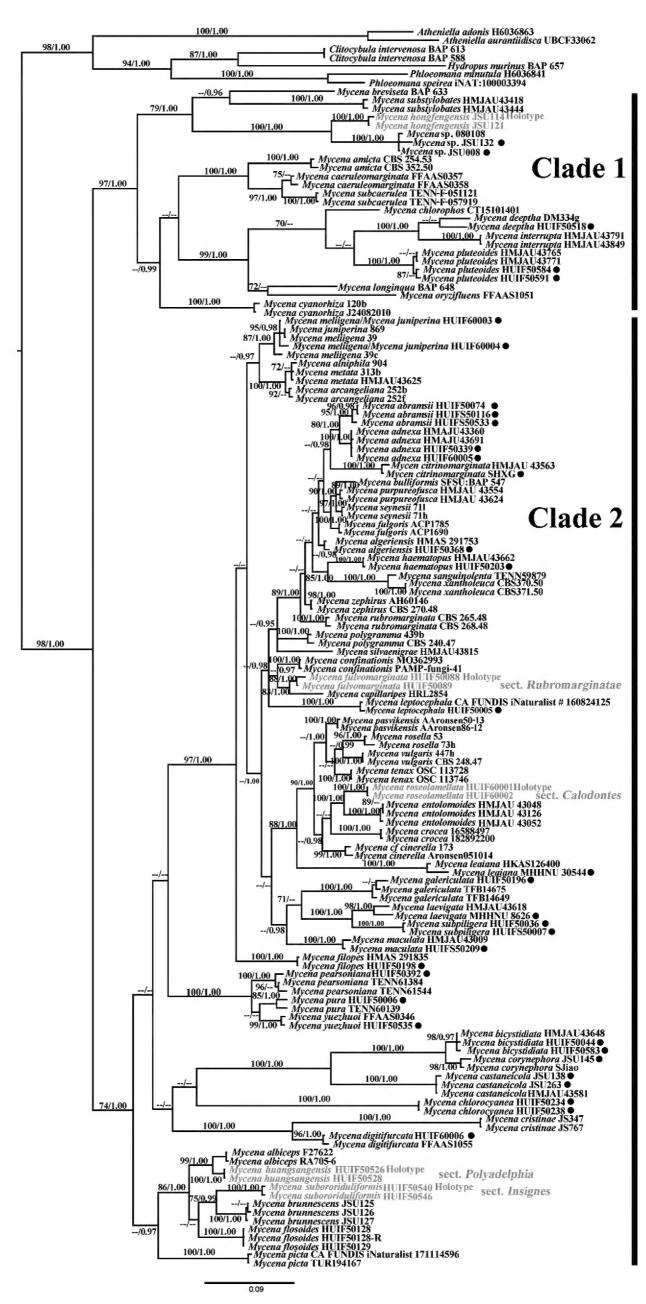


Figure 1. Phylogenetic relationships of *Mycena* species in Hunan Province inferred from the combined dataset (ITS and nrLSU) using Bayesian posterior probabilities (BP)  $\geq$  0.95; Bootstrap support (BS)  $\geq$  70% are reported on the branches. Red text represents new taxa. The black dots indicate the *Mycena* species collected from Hunan Province.

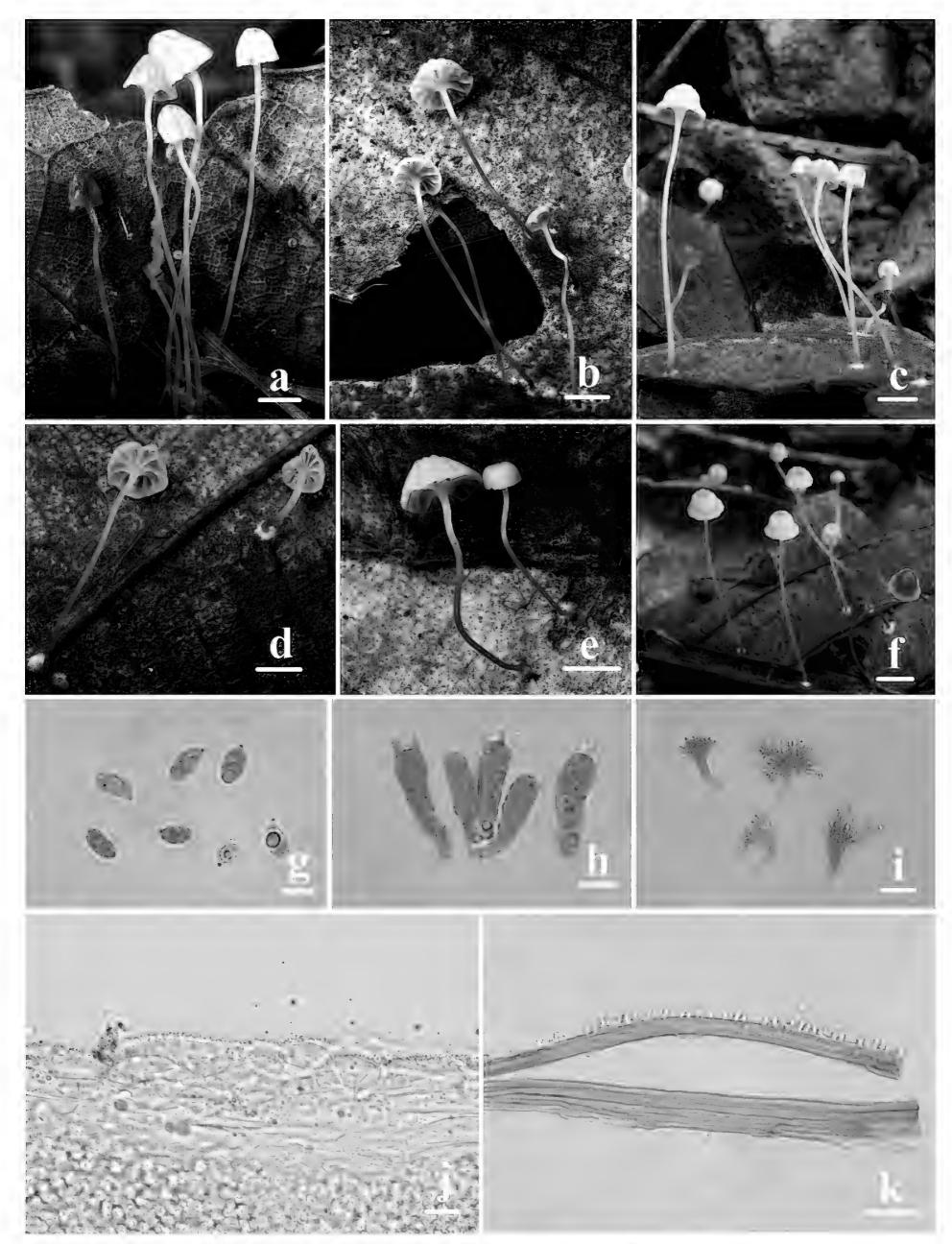


Figure 2. Basidiomata and microscopic features of *Mycena huangsangensis*  $\mathbf{a}$ – $\mathbf{f}$  basidiomata  $\mathbf{g}$  basidiospores  $\mathbf{h}$  basidia  $\mathbf{i}$  cheilocystidia  $\mathbf{j}$  pileipellis  $\mathbf{k}$  stipitipellis. Structures ( $\mathbf{g}$ – $\mathbf{i}$ ,  $\mathbf{k}$ ) were stained in 1% Congo red solution and  $\mathbf{j}$  were rehydrated in 5% KOH solution. Scale bars: 5 mm ( $\mathbf{a}$ – $\mathbf{f}$ ); 5  $\mu$ m ( $\mathbf{g}$ ); 10  $\mu$ m ( $\mathbf{h}$ – $\mathbf{k}$ ).

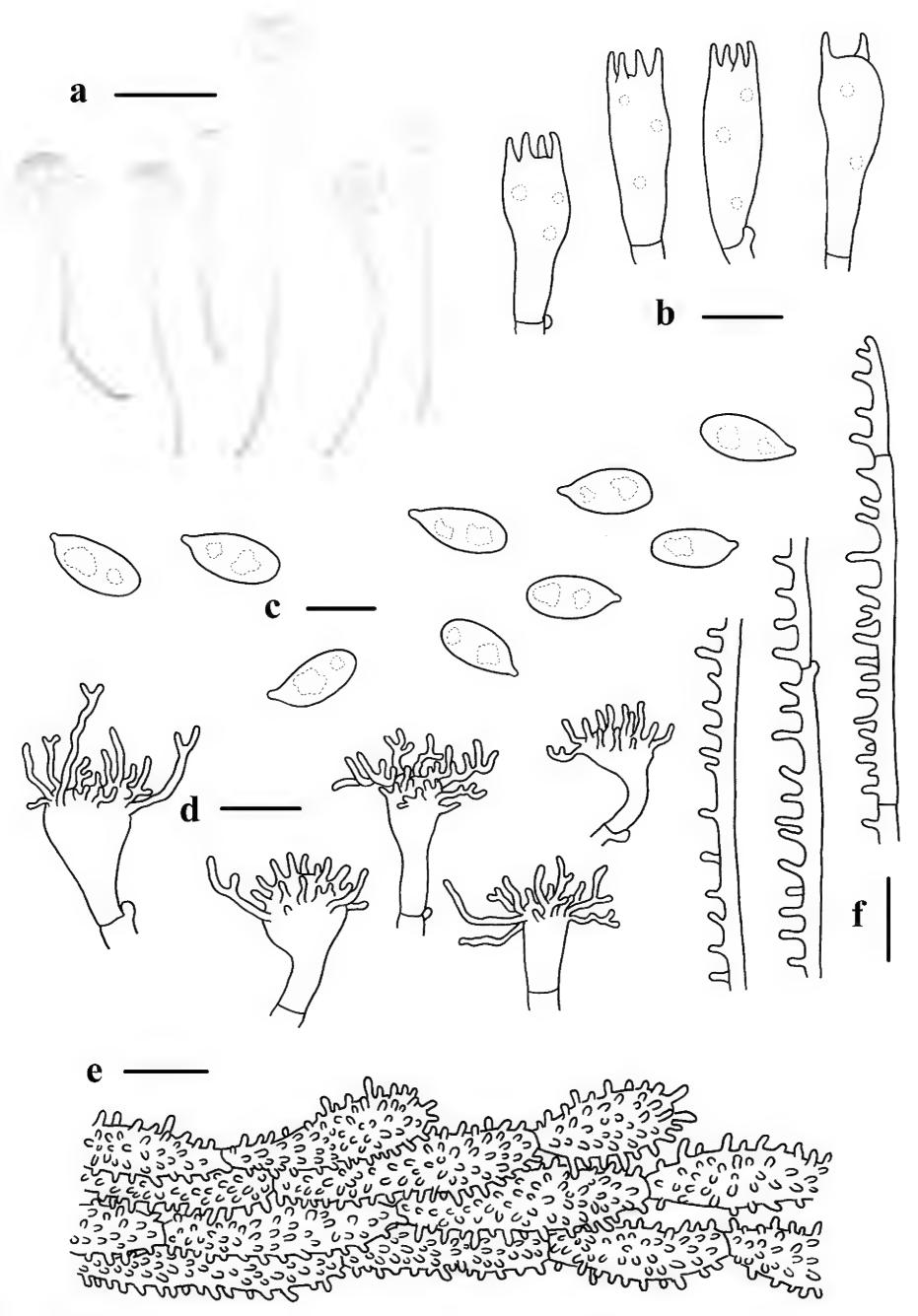


Figure 3. Morphological features of *Mycena huangsangensis* **a** basidiomata **b** basidia **c** basidiospores **d** cheilocystidia **e** pileipellis **f** hyphae of stipitipellis. Scale bars: 1 cm (**a**); 5 μm (**c**); 10 μm (**b**, **d**–**f**).

**Description.** Pileus 1–5 mm diam., hemispherical to obtusely conical, expanding with age, umbilicate or depressed center, sulcate, translucent-striate, pruinose and pubescent, light brown (6B4–6B7) to dark brown (6E7–6E8), or brownish-pink (7A4–7B4), paler brown towards margin. Context white, thin, fragile. Lamellae 9–11 reach the stipe, with 1–2 tiers of lamellulae, decurrent, white (4A1) to brown (7D7), edge concolorous with face. Stipe 6–25 × 1–2 mm, cylindrical, hollow, fragile, light brown (6B4–6B7) to brown (6E5–6E8) at the base, gradually becoming paler to white (4A1) towards the apex. Base covered with white fibrils. Odor and taste indistinctive.

Basidiospores  $6.0-7.4~(7.8)\times(3.2)~3.4-4.2~(4.3)~\mu\text{m}$ , Q = 1.4-2.1, Q =  $1.8\pm0.2$ , pip-shaped, elongated, hyaline, smooth, thin-walled, amyloid. Basidia  $15.4-20.9\times6.2-9.0~\mu\text{m}$ , 4-spored, clavate. Cheilocystidia  $13.2-25.0\times8.5-19.9~\mu\text{m}$ , abundant, clavate to obpyriform, covered with fairly numerous, simple to furcate, cylindrical excrescences.  $1.2-9.5\times0.5-1.4~\mu\text{m}$ . Pleurocystidia absent. Hyphae of the pileipellis  $12-27~\mu\text{m}$  wide, densely covered with warts or short cylindrical excrescences. Hyphae of the stipitipellis  $1.0-3.0~\mu\text{m}$  wide, densely covered with simple, cylindrical excrescences  $1.0-3.2\times0.8-1.5~\mu\text{m}$ . Clamp connections are present in the basidia, pileipellis, and stipitipellis hyphae.

Habitat. Gregarious on decaying leaves of deciduous trees.

Known distribution. Shaoyang City, Hunan Province.

**Additional materials examined.** CHINA• Hunan Province, Shaoyang City, Suining County, Hunan Huangsang National Nature Reserve, 26°24'21"N, 110°05'36"E, elev. 675 m, 24 April 2024, LiNa Liu, *HUIF50528* (collection number NN528).

Notes. Mycena huangsangensis can be considered to be a member of sect. Polyadelphia owing to very small basidiomata, a small number of lamellae, and a slender stipe and hyphae of the pileipellis, which are ornamented with short warts. Mycena huangsangensis belongs to the section with a brownish pileus, while M. alniphila Robich shows the most significant morphological similarity to M. huangsangensis. They have similar basidiomata color and shape of cheilocystidia, pileipellis hyphae densely covered with cylindrical excrescences, and diverticulate stipitipellis hyphae. However, M. alniphila differs in having adnate lamellae, slightly longer spores measuring  $8.5-11.0 \times (3.5) \cdot 4.0-5.5 \mu m$ , simple cheilocystidia without branching, and caulocystidia present (Robich 2003). Mycena albiceps and M. catalaunica Robich are somewhat similar to the new species; in particular, they share the same basidiomata shape and similar habitats. Mycena albiceps differs in the white colors of the pileus and black stipe (Gilliam 1976; Maas Geesteranus 1986). The latter, M. catalaunica, has a pale violaceous pink to pale vinaceous pink pileus, subglobose spores, and caulocystidia; the clamp connection is absent in all tissues, and cheilocystidia are subglobose (Robich 2003).

# Mycena fulvomarginata L.N. Liu, sp. nov.

MycoBank No: 856027

Figs 4, 5

**Diagnosis.** Differs from the closest species, *M. rubromarginata*, in having yellow lamellae edges and light-yellow contents in cheilocystidia, hyphae of the pileipellis, and stipitipellis.

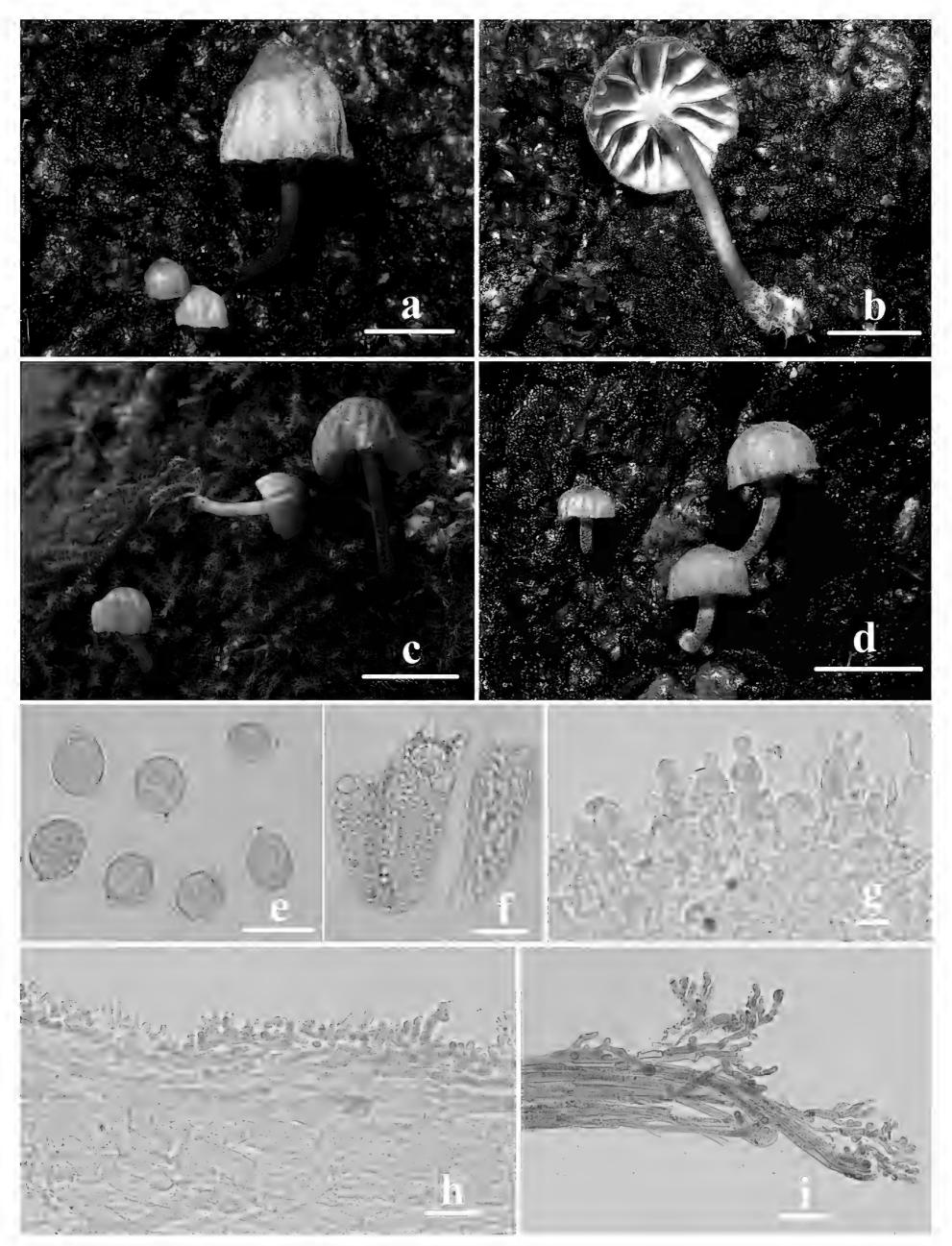


Figure 4. Basidiomata and microscopic features of *Mycena fulvomarginata* **a**–**d** basidiomata **e** basidiospores **f** basidia and basidioles **g** cheilocystidia **h** pileipellis **i** stipitipellis. Structures (**e**, **f**, **i**) were stained in a 1% Congo red solution, and **g**, **h** were rehydrated in a 5% KOH solution. Scale bars: 5 mm (**a**–**d**); 10 μm (**e**–**i**).

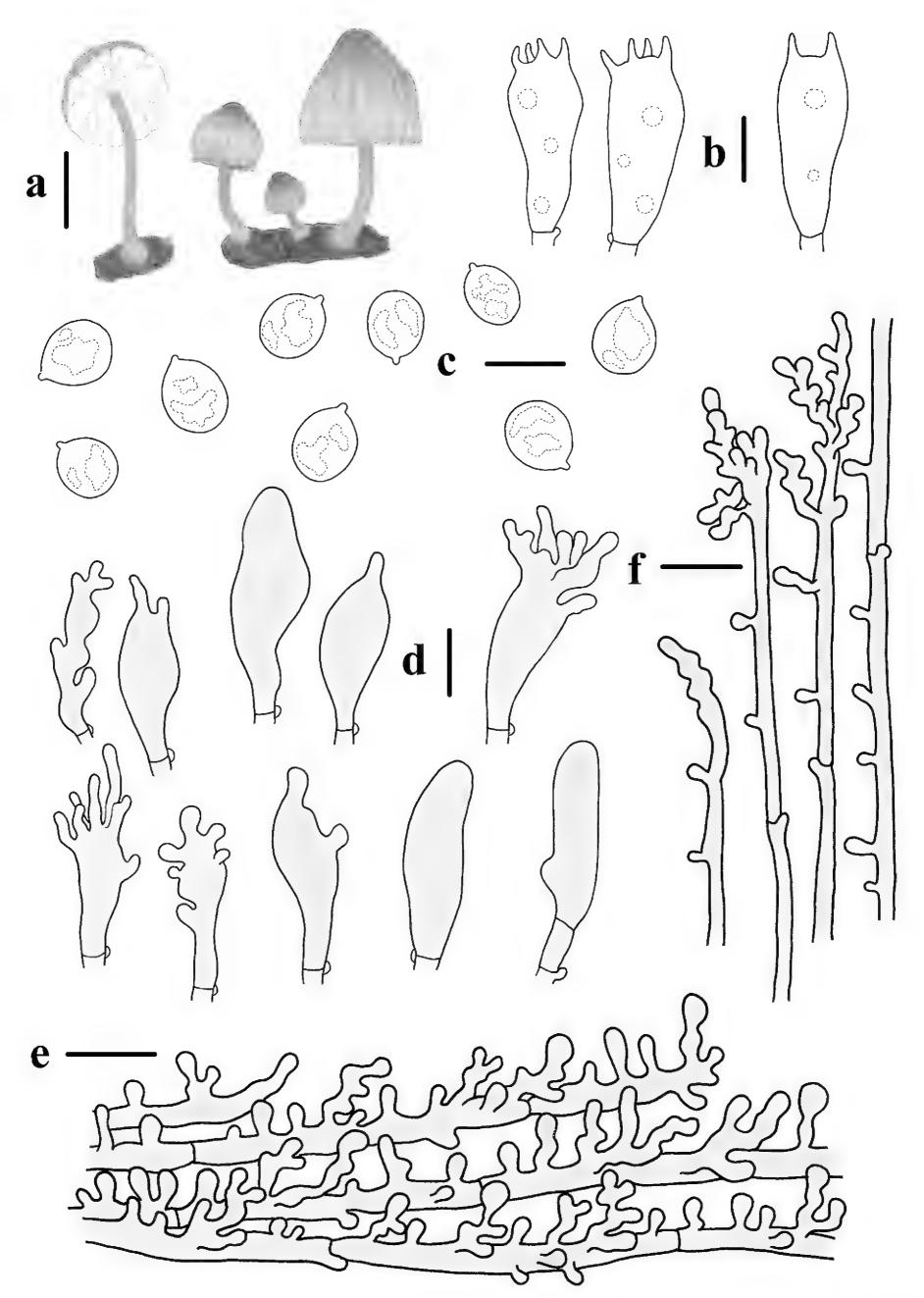


Figure 5. Morphological features of *Mycena fulvomarginata* **a** basidiomata **b** basidia **c** basidiospores **d** cheilocystidia **e** pileipellis **f** hyphae of stipitipellis. Scale bars: 5 mm (a); 10  $\mu$ m (b-f).

**Holotype.** CHINA• Hunan Province, Suining County, Hunan Huangsang National Nature Reserve, Shaoyang City, 26°25'41"N, 110°03'27"E, elev. 1075 m, 26 June 2021, LiNa Liu, *HUIF50088* (collection number NN88).

Etymology. Refers to the yellow color of the lamellae edges.

**Description.** Pileus 4–8 mm diam., hemispherical when young, paraboloid or campanulate with age, sulcate, pellucid-striate, pruinose, apex with obtuse umbo, the margin infrequently out of flatness, dark reddish brown (7C8) at center, gradually becoming paler towards the margin to light brown (7A6), turning purple (12B5) with age. Context white, thin, fragile. Lamellae 10-12 reach the stipe, with 1 tier of lamellulae, adnexed, white (4A1), edge yellow (5B7), stipe  $6.0-12.0\times0.5-1.0$  mm, central, cylindrical, hollow, fragile, finely white pruinose and pubescent, pale brown (6D7) to brown (6F7), fading to purple (12B5). Base slightly bulbous, covered with white fibrils. Odor and taste indistinctive.

Basidiospores (7.8) 7.9–9.9 (10.0) × (4.5) 5.7–8.0 (8.2)  $\mu$ m, Q = 1.2–1.7, Q = 1.3 ± 0.1, subglobose to broadly ellipsoid, hyaline, smooth, thin-walled, amyloid. Basidia 24.8–33.5 × 8.2–13.5  $\mu$ m, 4-spored, clavate. Cheilocystidia 22.6–45.8 × 7.7–17.8  $\mu$ m, abundant, fusiform to ventricose, clavate, subcylindrical, or somewhat irregularly shaped, smooth or covered with one or more apical simple to furcate excrescences, with light yellow contents. Pleurocystidia absent. Hyphae of the pileipellis 3.0–9.0  $\mu$ m wide, covered with simple to much-branched excrescences, 2.0–6.0 × 2.0–4.0  $\mu$ m, with light yellow contents. Hyphae of the stipitipellis 2.0–5.0  $\mu$ m wide, covered with numerous simple to furcate cylindrical excrescences, 2.0–11.0 × 2.0–8.0  $\mu$ m, with light yellow contents. Clamp connections are present in the basidia, pileipellis, and stipitipellis hyphae.

**Habitat.** Gregarious on moss-covered bark of living deciduous trees.

**Known distribution.** Hunan Province and Guangxi Zhuang Autonomous Region, China.

**Additional materials examined.** CHINA • Guangxi Zhuang Autonomous Region, Baise City, Leye County, Yachang Orchid National Nature Reserve, 24°46'29"N, 106°20'09"E, elev. 1080 m, 24 April 2024, LiNa Liu, *HUIF50089* (collection number NN89).

**Notes.** Mycena fulvomarginata belongs to sect. Rubromarginatae Singer ex. Maas Geest. due to the very small basidiomata, yellow lamellar edges, cheilocystidia with colored contents, diverticular pileipellis, and stipitipellis hyphae, along with exhibiting a growth habit on decaying wood. Mycena fulvomarginata is similar to the species described in this section: M. rubromarginata (Fr.) P. Kumm., M. seynii Quél., and M. bulliformis B.A. Perry & Desjardin. They resemble M. fulvomarginata in the shape of their cheilocystidia, are covered with excrescences on the pileipellis and stipe cortical hyphae, and have a similar brown pileus. However, M. rubromarginata differs in that it has longer pileipellis excrescences, up to 36 μm, and cheilocystidia that are 20-65 (up to 90) μm long, with reddish-brown contents (Robich 2003; Aronsen and Læssøe 2016). Mycena seynii should be easy to separate from the new species by its medium basidiomata, larger basidiospores measuring 10.5-15.0 × 6.0-7.5 µm, reddish-purple edge, larger cheilocystidia measuring 30-85 × 8-16 μm, pileipellis, and stipitipellis hyphae with brown to dark red granular contents (Robich 2003; Aronsen and Læssøe 2016). In addition, M. bulliformis differs from M. fulvomarginata by its violet to violet-brown edges, ellipsoid to broadly ellipsoid spores, and some smooth pileipellis hyphae (Perry and Desjardin 2016).

# Mycena hongfengensis L.N. Liu, sp. nov.

MycoBank No: 856029

Figs 6, 7

**Diagnosis.** Differs from *M. castaneicola* in having smooth cheilocystidia, dermatocysts present in the pileipellis, and stipitipellis hyphae.

**Holotype.** CHINA • Hunan Province, Xiangxi Tujia-Miao Autonomous Prefecture, Jishou City, Hongfeng Forest Park, 28°16'23"N, 109°40'45"E, elev. 230 m, 22 April 2024, ZhuXiang Liu, *JSU114* (collection number JD114).

**Etymology.** Refers to the Hongfeng Forest Park, from where the holotype was collected.

**Description.** Pileus 2–5 mm diam., hemispherical when young, becoming nearly campanulate or plano-convex with age, with a centrally flattened depression, margin smooth, sulcate, translucently striate, pure white (4A1), white pubescent. Context pure white, thin, fragile. Lamellae 16–18 reach the stipe, with 1–2 tiers of lamellulae, narrowly free, pure white (4A1), concolorous with the sides. Stipe  $15-40\times0.1-0.5$  mm, almost equal or slightly expanding towards the base, hollow, white (4A1) to greyish-white (4B1), pubescent or puberulous, base swollen. With a not well-developed basal disc, covered with white hirsute. Odor and taste not distinctive.

Basidiospores (6.2) 6.3–7.6 (7.7) × (3.4) 3.5–4.9 (5.2) µm, Q = 1.4–2.0, Qm = 1.7  $\pm$  0.2, oblong or pip-shaped, hyaline, thin-walled, amyloid. Basidia 8–16 × 4–8 µm, two- and four-spored, clavate, hyaline. Cheilocystidia 11–43 × 5–9 µm, obpyriform, fusiform, ventricose, filiform, with a long neck, up to 25 µm, with an acute and occasionally branched apex, hyaline. Pleurocystidia absent. Pileipellis hyphae 2–13 µm wide, hyphae cylindrical, densely covered with warts and cylindrical excrescences,  $1.0-6.0 \times 1.0-2.0$  µm, with irregularly cylindric to strangulated dermatocysts,  $63-200 \times 7-20$  µm, walls 1.0-2.0 µm, greenish grey (1C2). Hyphae of the stipitipellis 1.0-6.0 µm wide, smooth, dermatocysts numerous, clavate to pyriform,  $50-320 \times 5-20$  µm, long, flexuous, filiform, simple, and tapering towards the apex. Clamp connections are absent in the basidia, pileipellis, and stipitipellis hyphae.

**Habitat.** Gregarious on decaying leaves of deciduous trees.

**Known distribution.** Xiangxi Tujia-Miao Autonomous Prefecture, Hunan Province.

Additional materials examined. CHINA• Hunan Province, Xiangxi Tujia-Miao Autonomous Prefecture, Jishou City, Hongfeng Forest Park, 28°16'26"N, 109°40'48"E, elev. 255 m, 22 April 2024, ZhuXiang Liu, *JSU121* (collection number JD121).

**Notes.** *Mycena hongfengensis* is characterized by its pure white basidiomata, free lamellae, oblong to pip-shaped spores, and the presence of dermatocysts in the pileipellis and stipitipellis. According to the Maas Geesteranus classification, the new species could belong to an uncertain position. When we first found this specimen in the field, we thought it might be a member of either sect. *Saccharifera* or sect. *Amparoina*. All species in two sections have a white basidiomata, pubescent pileus and stipe, and stipe with a basic disc (Maas Geesteranus 1983; Desjardin 1995; Na and Bau 2019b). However, *M. hongfengensis* can be clearly distinguished from other species through microscopic characteristics. The presence of dermatocysts in the pileipellis and stipitipellis is the most important

characteristic for separating *M. hongfengensis* from species of sect. *Sacchariferae* and sect. *Amparoina. Mycena castaneicola* T. Bau & Q. Na is the most similar to the new species in the macroscopic characteristics, but the microscopic features differ significantly. Sequences labeled as *Mycena* sp., which were extracted from

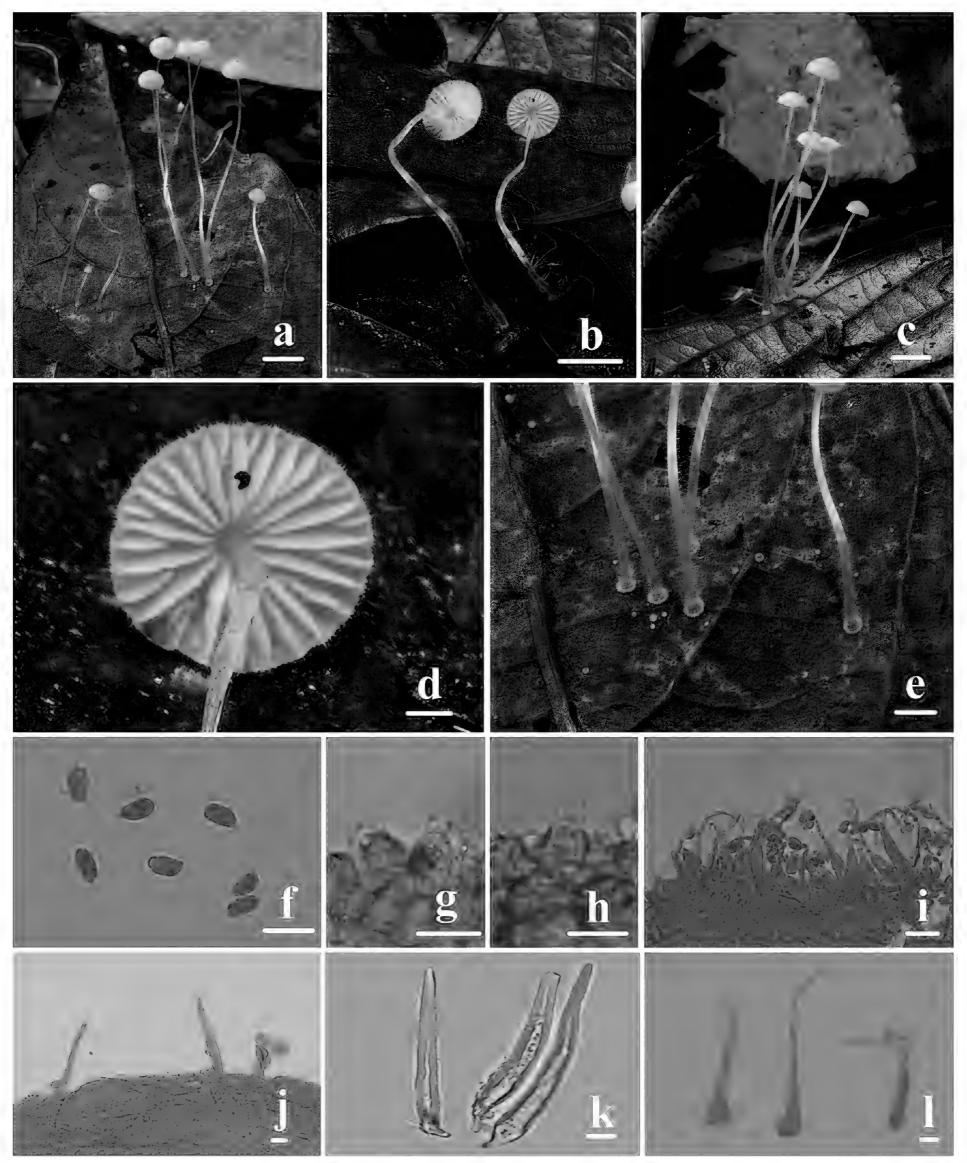


Figure 6. Basidiomata and microscopic features of *Mycena hongfengensis*  $\mathbf{a}-\mathbf{e}$  basidiomata  $\mathbf{f}$  basidiospores  $\mathbf{g}$ ,  $\mathbf{h}$  basidia  $\mathbf{i}$  cheilocystidia  $\mathbf{j}$ ,  $\mathbf{k}$  dermatocysts in the pileipellis  $\mathbf{l}$  dermatocysts in the stipitipellis. Structures  $(\mathbf{f}-\mathbf{j}, \mathbf{l})$  were stained in a 1% Congo red solution and  $\mathbf{k}$  were rehydrated in a 5% KOH solution. Scale bars: 5 mm  $(\mathbf{a}-\mathbf{c}, \mathbf{e})$ ; 1 mm  $(\mathbf{d})$ ; 5  $\mu$ m  $(\mathbf{f})$ ; 10  $\mu$ m  $(\mathbf{g}-\mathbf{l})$ .

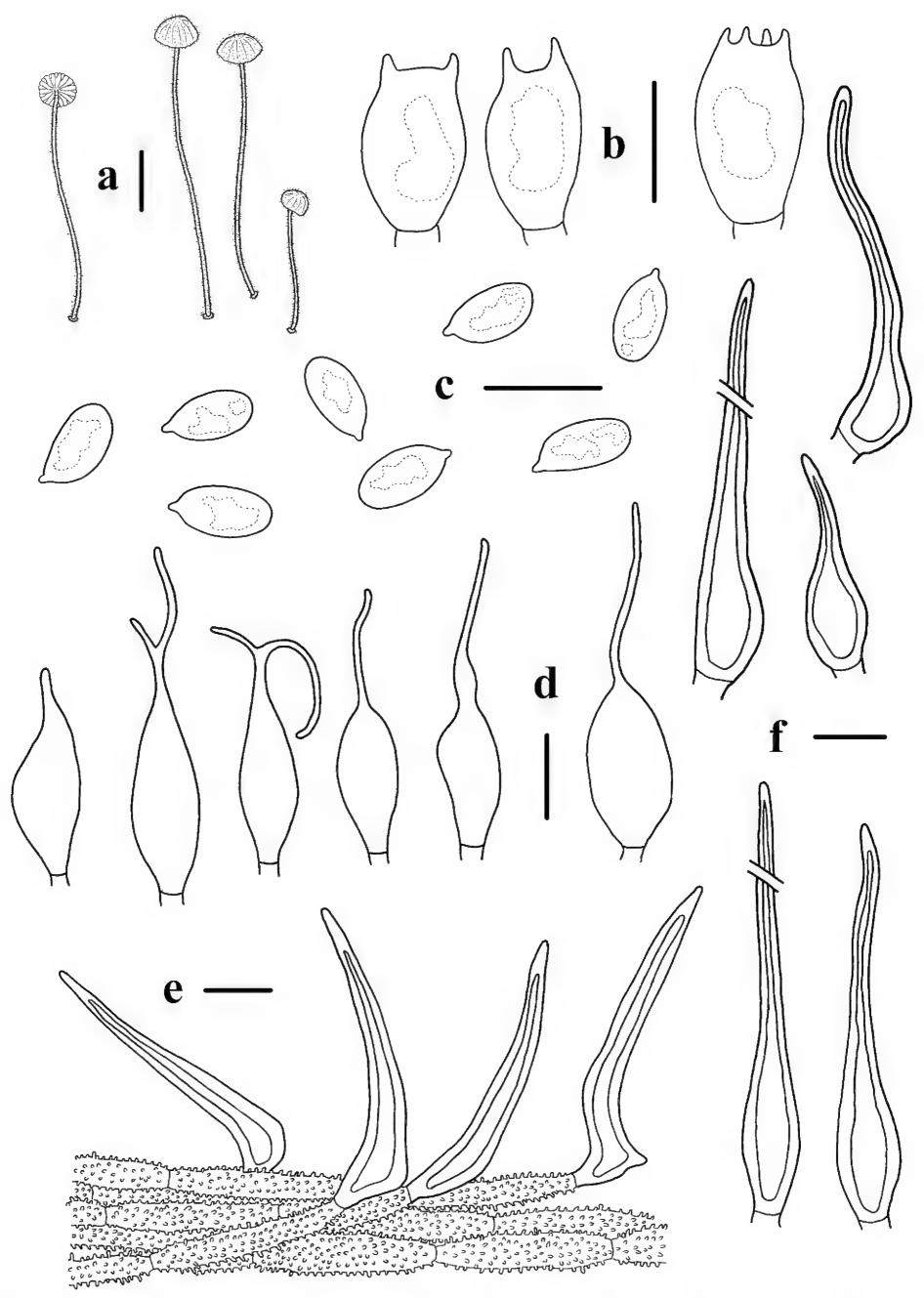


Figure 7. Morphological features of *Mycena hongfengensis* **a** basidiomata **b** basidia **c** basidiospores **d** cheilocystidia **e** dermatocysts in the pileipellis **f** dermatocysts in the stipitipellis. Scale bars: 1 cm (**a**); 10  $\mu$ m (**b**-**f**).

the host *Myrica rubra* (Lour.) Siebold & Zucc. and originated from Japan, along with the sequence of *M. hongfengensis*, formed a well-supported lineage in the phylogenetic analysis (BS/BP = 100/1.00). We are very fortunate to have collected *Mycena* sp. in the field. However, we only collected one tiny basidioma; the microscopic structures we observed are not comprehensive. Both taxa have white pileus and pubescent pileus and stipe. The uncertain *Mycena* species differs in pileipellis, which has "acanthocysts" that are spherical or vesicular and covered with spines. These features can be used to easily differentiate the two species.

#### Mycena subroriduliformis L.N. Liu, sp. nov.

MycoBank No: 856045

Figs 8, 9

**Diagnosis.** Differs from *M. surculosa* in having a viscid pileus and stipe.

**Holotype.** CHINA • Hunan Province, Suining County, Hunan Huangsang National Nature Reserve, Shaoyang City, 26°24'47"N, 110°05'20"E, elev. 542 m, 25 April 2024, LiNa Liu, *HUIF50540* (collection number NN540).

Etymology. Refers to the viscid pileus and stipe of the new species.

**Description.** Pileus 2–8 mm diam., hemispherical when young, campanulate with age, with obvious depression at the center, sulcate, translucent-striate, surface wet, glabrous, brownish white (7A1), light brown (7C3), brownish grey (7D1–6D2) when old. Context white, thin, and fragile. Lamellae 18-20 reach the stipe, with 1-2 tiers of lamellulae, decurrent, white (4A1), concolorous with the sides. Stipe  $2-45 \times 1-2$  mm, cylindrical, hollow, fragile, surface glutinous, white (4A1) to brownish grey (5A1–5D3) towards the apex, light brown to brown (5D6–6D6) towards the base, base swollen. Odor and taste indistinctive.

Basidiospores (6.2) 6.6–8.5 (9.0) × (3.6) 3.8–5.2 (5.3) µm, Q = 1.5–2.1, Q = 1.7  $\pm$  0.1, pip-shaped, cylindrical, hyaline, amyloid, smooth. Basidia 19.3–26.9 × 5.6–8.0 µm, 4-spored, clavate, hyaline. Cheilocystidia 16.8–26.9 × 6.3–17.1 µm, abundant and variably shaped, clavate to cylindrical with short, more or less branched projections, 1.0–6.0 × 1.0–2.0 µm, thin-walled, hyaline. Pleurocystidia absent. Pileipellis hyphae somewhat gelatinized, 2.0–5.0 µm wide, with irregular simple to branched warts or cylindrical excrescences, 1.0–4.0 × 1.0–2.0 µm. Hyphae of the stipitipellis 1.0–6.0 µm wide, covered with cylindrical excrescences. 1.0–4.0 × 1.0–2.0 µm. Clamp connections are present in the basidia, pileipellis, and stipitipellis hyphae.

Habitat. Gregarious on decaying leaves of deciduous trees.

Known distribution. Shaoyang City, Hunan Province.

**Additional materials examined.** CHINA• Hunan Province, Suining County, Hunan Huangsang National Nature Reserve, Shaoyang City, 26°24'39"N, 110°05'25"E, elev. 588 m, 25 April 2024, LiNa Liu, *HUIF50546* (collection number NN546).

**Notes.** The following characteristics placed this new species in the sect. *Insignes* Maas G. due to the viscid pileus and stipe, decurrent lamellae, pip-shaped spores, clavate cheilocystidia with coarse excrescences, hyphae of the pileipellis embedded in gelatinous matter, and diverticulate (Maas Geesteranus 1989). The three other similar species in sect. *Insignes* are *M. surculosa* Maas G. & de Meijer, *M. odorifera* (Peck) Sacc., and *M. calceata* Robich. All are differentiated from *M. subroriduliformis* as follows: *M. surculosa* has a dry pileus, with only

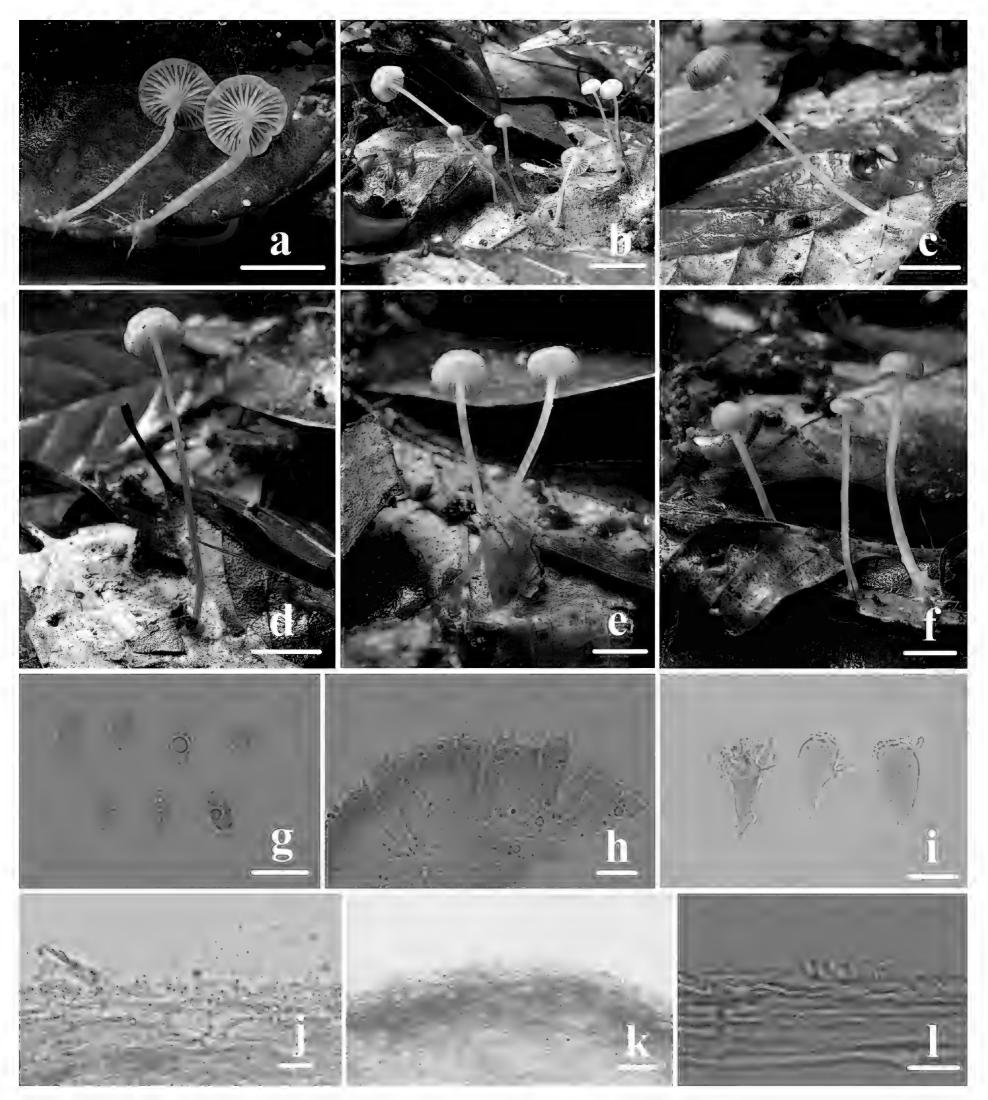


Figure 8. Basidiomata and microscopic features of *Mycena subroriduliformis*  $\mathbf{a}$ – $\mathbf{f}$  basidiomata  $\mathbf{g}$  basidiospores  $\mathbf{h}$  basidia  $\mathbf{i}$  cheilocystidia  $\mathbf{j}$ ,  $\mathbf{k}$  pileipellis  $\mathbf{l}$  stipitipellis. Structures ( $\mathbf{g}$ – $\mathbf{i}$ ,  $\mathbf{k}$ ,  $\mathbf{l}$ ) were stained in a 1% Congo red solution and  $\mathbf{j}$  were rehydrated in a 5% KOH solution. Scale bars: 1 cm ( $\mathbf{a}$ – $\mathbf{f}$ ); 10  $\mu$ m ( $\mathbf{g}$ – $\mathbf{l}$ ).

the stipe being viscid. The apical side branches of the pileipellis are densely covered and simple to forked, with cylindrical excrescences. The stipitipellis is somewhat gelatinized (Maas Geesteranus and De Meijer 1997). *Mycena odorifera* has a distinctive alkaline-like odor, a pruinose stipe, pubescence, and cheilocystidia occasionally with forked apices (Smith 1935). *Mycena calceata* has dark brown basidiomata and relatively large spores (11.0–13.5  $\times$  5.5–8.0  $\mu$ m), smooth cheilocystidia, or one or two branches at the apex (Robich 2003).

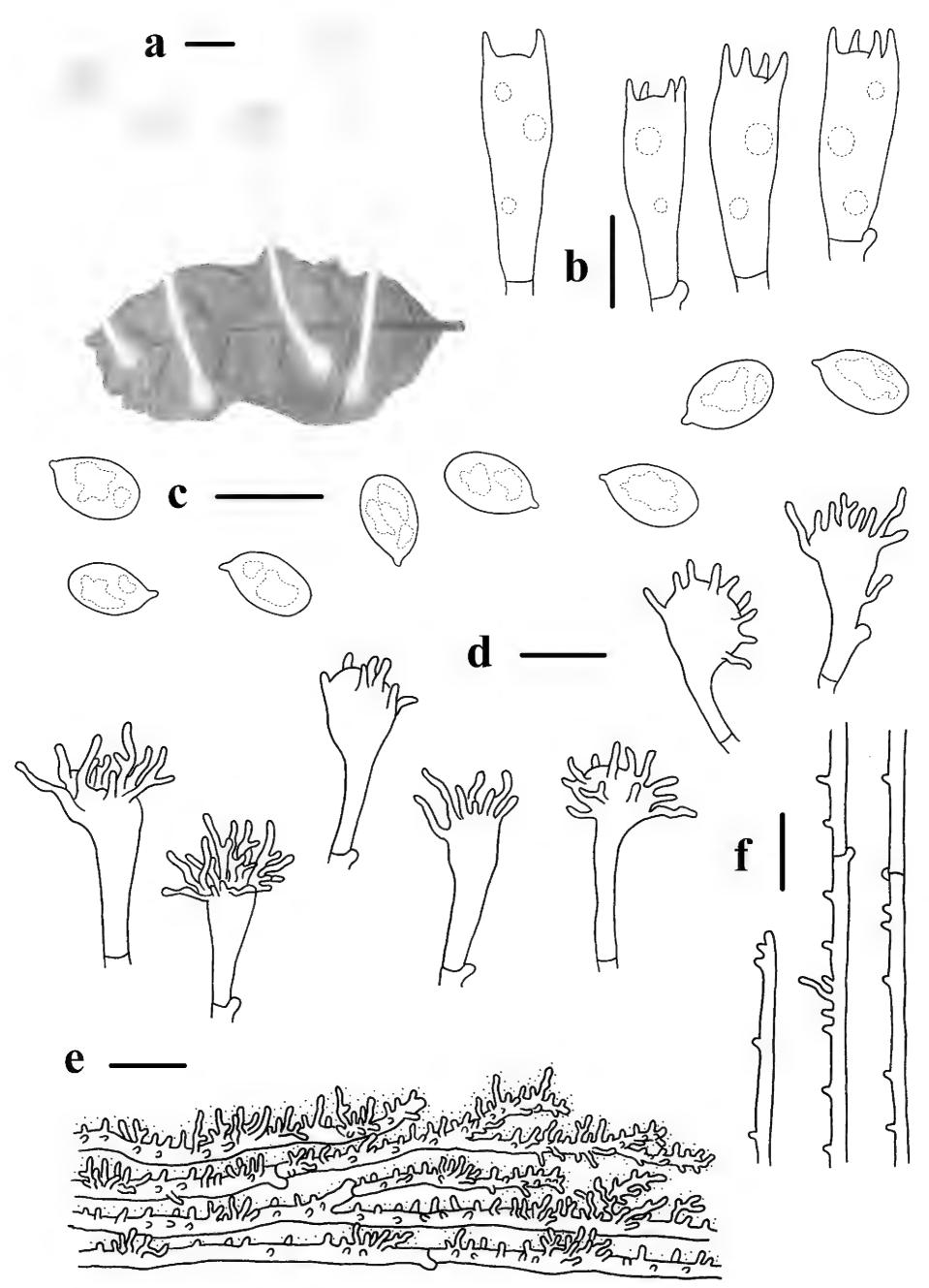


Figure 9. Morphological features of *Mycena subroriduliformis* **a** basidiomata **b** basidia **c** basidiospores **d** cheilocystidia **e** pileipellis **f** hyphae of stipitipellis. Scale bars: 5 mm (**a**); 10 μm (**b**–**f**).

# Mycena roseolamellata L.N. Liu, sp. nov.

MycoBank No: 856026

Figs 10, 11

**Diagnosis.** Differs from *M. pura* in having a brown pileus and pink lamellae.

**Holotype.** CHINA• Hunan Province, Ningxiang City, Biandan'ao, Lijingpu Subdistrict, 28°12'07"N, 112°32'43"E, elev. 110 m, 28 November 2023, ShengQiang Liu, *HUIF60001* (collection number NN601).

**Etymology.** Refers to the pink colors of lamellae.

**Description.** Pileus 7–17 mm diam., parabolic when young, then campanulate or broadly conical with age, apex with an obtuse umbo, sulcate, translucent-striate, glabrous, dark brown (6F5–6F8) at first, then turning pale brownish yellow (6A4) to pale brown (6E4) with age, margin brownish white (6A2) to pale brown (6D4–6D6). Context white, fragile, thin. Lamellae 24–26 reach the stipe, with 1–2 tiers of lamellulae, adnate or slightly adnex, white (4A1) when young, pinkish to light pink (9A3–9A2) at maturity, concolorous with faces. Stipe 29–30  $\times$  2.0–3.0 mm, central, cylindrical, hollow, dark brown (6F5-6F8), pale brownish yellow (6A4) to pale brown (6D7) with age, base covered with long, dense, white fibrils. Odor and taste indistinctive.

Basidiospores (8.3) 8.6–10.8 (11.5) × (5.3) 5.4–6.3 (6.4)  $\mu$ m, Q = 1.5–2.0, Q = 1.7 ± 0.1, ellipsoid to elongated, hyaline, smooth, thin-walled, amyloid. Basidia 21.7–30.8 × 6.8–8.9  $\mu$ m, 2-spored, clavate. Cheilocystidia 26.7–84.9 × 8.6–18.7  $\mu$ m, abundant, fusiform, ventricose-rostrate, obtuse apex, base tapered, with short to long stalk, smooth, hyaline, amyloid, thin-walled. Pleurocystidia similar to cheilocystidia, 52.3–105.7 × 12.3–20.9  $\mu$ m. Pileipellis 1.0–6.0  $\mu$ m wide, smooth, terminal hyphae sometimes diverticulate, 1.0–6.0 × 1.0–2.0  $\mu$ m. Stipitipellis 2.0–5.0  $\mu$ m, smooth, terminal hyphae sometimes diverticulate, 1.0–4.0 × 1.0–2.0  $\mu$ m. Clamp connections are absent in the basidia, pileipellis, and stipitipellis hyphae.

**Habitat.** Gregarious on decayed twigs of bamboo or woody debris of deciduous trees.

Known distribution. Ningxiang City, Hunan Province.

**Additional materials examined.** CHINA • Hunan Province, Ningxiang City, Biandan'ao, Lijingpu Subdistrict, 28°12'07"N, 112°32'43"E, elev. 110 m, 22 December 2023, ShengQiang Liu, *HUIF60002* (collection number NN602).

**Notes.** *Mycena roseolamellata* is classified into sect. *Calodontes* based on the smooth cheilocystidia and stipitipellis. Microscopically, *M. pura* (Pers.) P. Kumm. is the most similar to *M. roseolamellata*; however, *M. pura* is distinguished from *M. roseolamellata* by its purple pileus with pinkish or brown tints, lamellae interveined with age, the presence of clamp connections in all tissues, and the absence of a root-like, pruinose stipe (Robich 2003; Aronsen and Læssøe 2016). *Mycena rosea* Gramberg is somewhat similar to *M. roseolamellata*; they have pink lamellae, smooth cheilocystidia, and pleurocystidia. *Mycena rosea* can be distinguished from *M. roseolamellata* by having a pink pileus with a dull yellow center, a white or pink stipe, and the presence of clamp connections in all tissues (Robich 2003; Aronsen and Læssøe 2016). *Mycena roseolamellata* is very different from any species of sect. *Calodontes*, owing to its brown pileus. *Mycena galericulata* (Scop.) Gray shares some similarities with *M. roseolamellata* in terms of pileus color, but *M. galericulata* is differentiated by the presence of non-smooth cheilocystidia, pileipellis, and stipitipellis (Maas Geesteranus 1992a, 1992b).

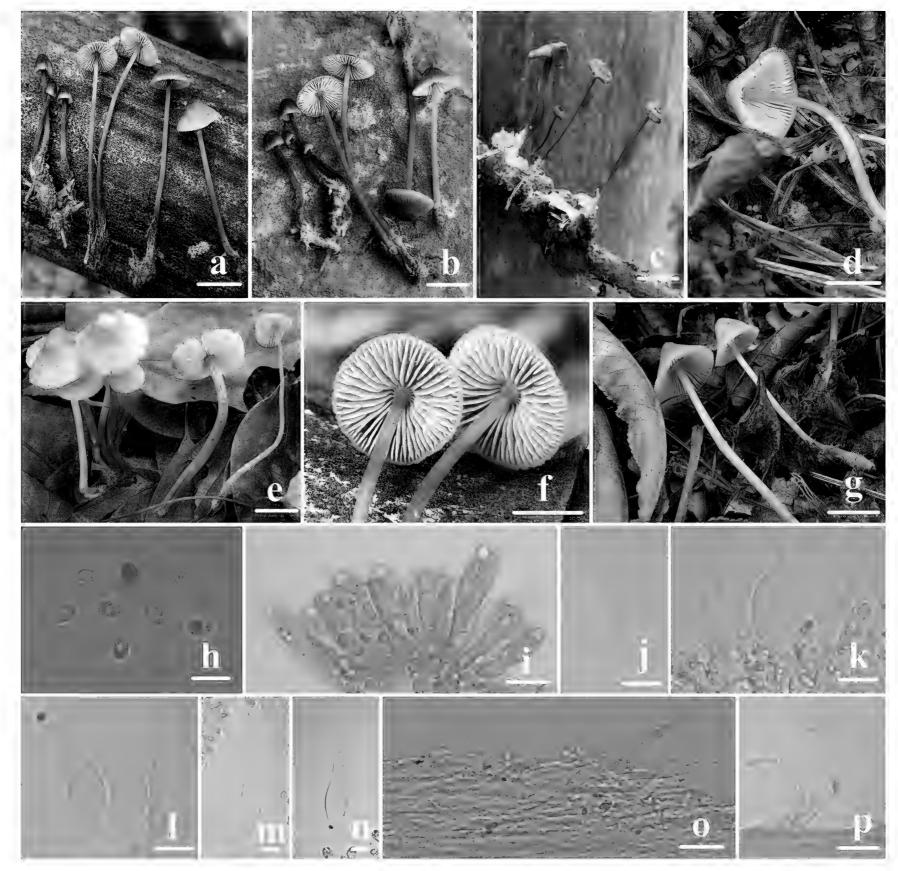


Figure 10. Basidiomata and microscopic features of *Mycena roseolamellata*  $\mathbf{a}-\mathbf{g}$  basidiomata  $\mathbf{h}$  basidiospores  $\mathbf{i}$ ,  $\mathbf{j}$  basidia  $\mathbf{k}$ ,  $\mathbf{l}$  cheilocystidia  $\mathbf{m}$ ,  $\mathbf{n}$  pleurocystidia  $\mathbf{o}$  pileipellis  $\mathbf{p}$  stipitipellis. Structures  $(\mathbf{j}, \mathbf{p})$  were stained in 1% Congo red solution, and  $\mathbf{h}$ ,  $\mathbf{i}$ ,  $\mathbf{k}-\mathbf{o}$  were rehydrated in 5% KOH solution. Scale bars: 1 cm  $(\mathbf{a}-\mathbf{g})$ ; 10  $\mu$ m  $(\mathbf{h}-\mathbf{p})$ .

# **Discussion**

The combination of morphological features and phylogenetic analyses revealed the presence of 30 species in Hunan Province, including five new species. In the Maas Geesteranus classification, the new species *M. fulvomarginata*, *M. huangsangensis*, *M. subroriduliformis*, and *M. roseolamellata* are classified into sect. *Rubromarginatae*, sect. *Polyadelphia*, sect. *Insignes*, and sect. *Calodontes*, respectively (Maas Geesteranus 1980, 1992a, 1992b; Maas Geesteranus and De Meijer 1997; Robich 2003, 2016). However, we could not place the new species *M. hongfengensis* in any section at present due to its special characteristics. The taxonomy of *Mycena* is overly complex, and infrageneric classification generally relies on the reported morphology of *Mycena*, and the characteristics of some species are not consistent with the common features shared by species of the numerous sections. Therefore, we need to increase the number of related species to identify the common characteristics of those species and further improve the taxonomy of *Mycena*.

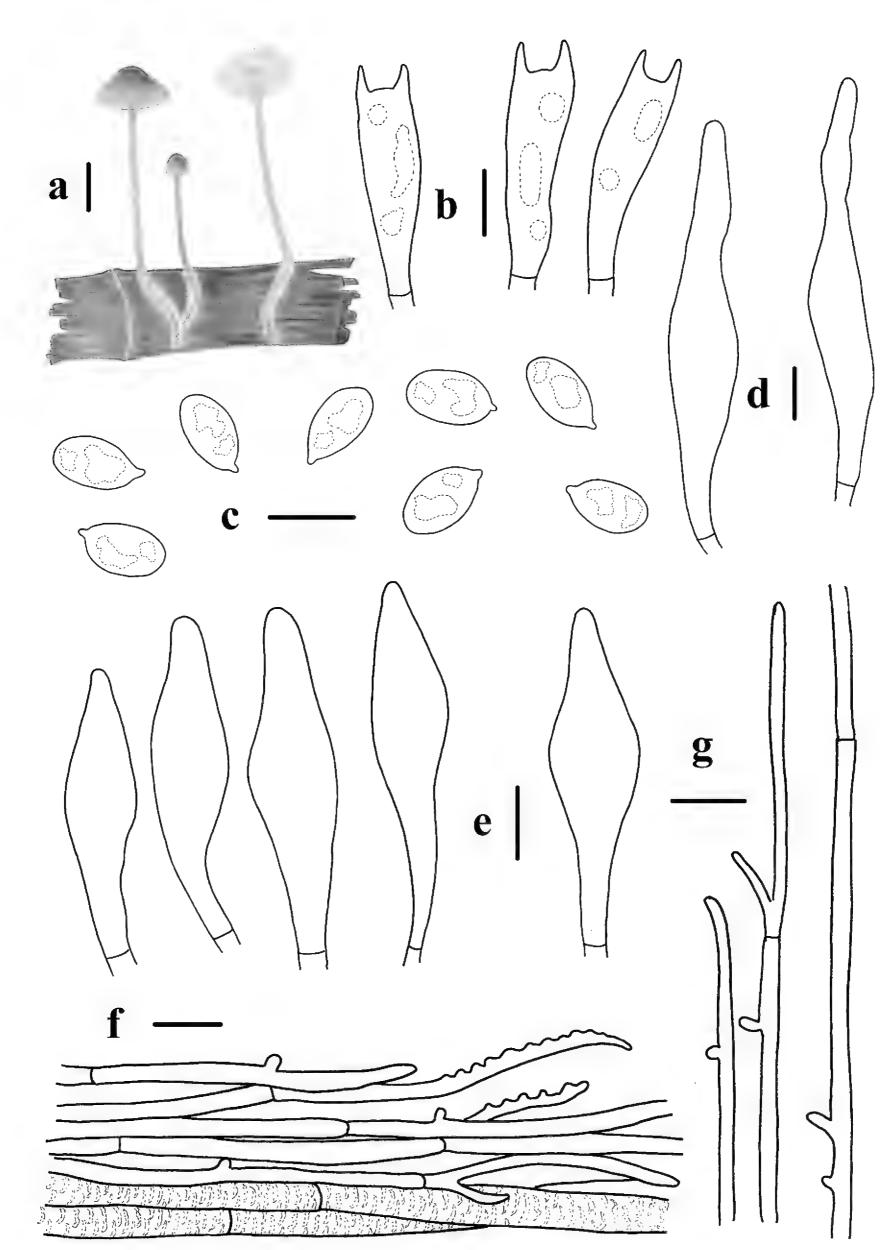


Figure 11. Morphological features of *Mycena roseolamellata* **a** basidiomata **b** basidia **c** basidiospores **d** pleurocystidia **e** cheilocystidia **f** pileipellis **g** hyphae of stipitipellis. Scale bars: 1 cm (a); 10  $\mu$ m (b-g).

Among the 30 *Mycena* species collected in Hunan, we found *M. picta* (Fr.) Harmaja on Yuelu Mountain, Hunan. As we obtained only one basidiomata in the field, sequences of *M. picta* could not be obtained for phylogenetic analysis.

However, we can confirm that the specimen is *M. picta* based on its distinct macroscopic and microscopic characteristics (Aronsen and Læssøe 2016; Bau et al. 2021). And *M. heteracantha* was also collected in Yuelu Mountain (Na and Bau 2019a). We are uncertain of the classification of *M. juniperina* Aronsen and *M. meliigena* (Berk. & Cooke) Sacc. Distinguishing the 8 related specimens we collected in Hunan was difficult because of their similar microcharacteristics and the limited number of sequences available for downloading from GenBank, although the color of their basidiomata varies. Therefore, we identified all the related specimens we collected as *M. juniperina/meliigena*, as we did not obtain type specimens. Additional studies are needed.

Among the phylogenetic trees, *M. hongfengensis*, *M. deeptha* Aravind. & Manim. and *M. pluteoides* T. Bau & Q. Na were grouped into clade 1. Interestingly, all species have a non-smooth pileipellis hyphae. Most species collected from Hunan are mainly concentrated in clade 2. Four new species are grouped in clade 2; all *Mycena* species in this clade have a stipe or a stipe base that is covered with white fibrils.

The distinct topography, climate, and flora of Hunan Province have produced abundant and distinctive *Mycena* specimens. We conducted a comparative analysis of the geographical compositions of the 30 *Mycena* species we collected and preliminarily divided the distribution of the genus *Mycena* in Hunan Province into the following four types (Table 1). The analysis of the floristic components of the species reveals that *Mycena* in Hunan Province is cosmopolitan, exhibiting tropical-subtropical floristic characteristics as well as a certain proportion of northern temperate characteristics, indicating that *Mycena* species in Hunan possess diverse and transitional features.

Although *Mycena* was widely distributed in the world, the earliest and most detailed research was conducted in Europe and North America (Singer and Digilio 1951; Robich 2003, 2016). The morphological and microscopic characteristics of the *Mycena* species collected in Hunan Province are basically consistent with those of materials from Europe. However, the sizes of the basidiomata, basidiospores, basidia, cheilocystidia, and caulocystidia occasionally vary subtly. This may be due to the geographical location and environmental changes in the species at that time. From a geographical point of view, the Mycena species we collected show some differences compared with those documented in monographs from Europe and North America. Free lamellae are important for diagnosing these species in Mycena. We found that Mycena with free lamellae are more commonly collected in Hunan Province than in temperate regions in China; for example, M. deeptha and M. pluteoides, belonging to sect. Exornatae Maas Geest, are widely distributed in the western and southern areas of Hunan. And members of the sect. Exornatae are most commonly found in subtropical regions of Asia (Desjardin et al. 2010; Aravindakshan et al. 2012; Aravindakshan and Manimohan 2013a; Bau et al. 2021). These findings also indirectly suggest that sect. Exornatae has a certain preference for geographical location. Our research may contribute to the exploration of the origin and evolution of Mycena.

# Key to the known species of Mycena in Hunan Province

1	Stipe arising from a basal disc	.2
_	Stipe not arising from a basal disc	.3

2	Dermatocysts present in the pileipellis and stipitipellis	
_	Dermatocysts absent in the pileipellis and stipitipellis	
3	Fresh and young basidiomata exude colored fluid when	
		M. haematopus
-	Basidiomata do not exude colored fluid when damaged	7
4	Pileus glabrous, viscid, white, with a pale brown center,	depressed at the
	center	
_	Pileus dry, pubescent, pure white, not depressed at cent	
5	Cheilocystidia vesiculose, smooth	M. deeptha
_	Cheilocystidia densely covered with projections	M. pluteoides
6	Basidiomata growing on Castanea burs, pileus slightly p	
-	Basidiomata growing on dead wood or humus layer, pil	
	covering	M. heteracantha
7	Lamellae not white, or occasionally white when young	
_	Lamellae white	
8	Lamellae faces pink, occasionally white when young, o	cheilocystidia hya-
	line	M. roseolamellata
-	Lamellae faces orange-yellow, cheilocystidia with yellow	v contents
9	Cheilocystidia smooth	
_	Cheilocystidia with simple to branched excrescences	
10	Lamellae faces not concolorous with the sides	
_	Lamellae faces concolorous with the sides	
11	Lamellae edges light yellow	
_	Lamellae edges light brown to yellowish-brown	M. fulvomarginata
12	Hyphae of the pileipellis smooth	
_	Hyphae of the pileipellis diverticulate	
13	Pileus brown	_
_	Pileus violet	
14	Pleurocystidia absent	
_	Pleurocystidia present	•
15	Pileipellis not gelatinized	•
-	Pileipellis gelatinized	M. pearsoniana
16	Lamellae adnate or adnexed	
_	Lamellae decurrent	
17	Pileus grey brown	
_	Pilues white	
18	Cheilocystidia thick-walled	
_	Cheilocystidia thin-walled	M. digitifurcata
19	Hyphae of the stipitipellis smooth, caulocystidia present	•
_	Hyphae of the stipitipellis covered with warty or divertical	
	absent	
20	Pileipellis and stipitipellis gelatinized	_
-	Pileipellis and stipitipellis not gelatinized	
21	Basidiomata sticky	
-	Basidiomata dry	
22	Pileus with yellow tone	
_	Pileus without vellow tone	24

ngth <i>M. picta</i>	Pileus bucket-shaped, lamellae broader than the le	23
ae broad	Pileus hemispherical, parabolical to convex, lamella	_
neliigena/M. juniperina		
	Basidiospores globose	24
25	Basidiospores broadly ellipsoid to ellipsoid	-
M. maculata	Pileus and lamella with red spots when old	25
26	Pileus and lamella without red spots when old	_
M. chlorocyanea	Rhizomorphs present	26
27	Rhizomorphs absent	_
M. galericulata	Clamp connections absent in all tissues	27
28	Clamp connections present in all tissues	_
pyriform to vesicular	Acanthocysts present, acanthocysts of two types,	28
M. bicystidiata		
	Acanthocysts absent	_
M. filopes	Pileus and stipe pruninose, iodoform when dry	29
M. huangsangensis	Pileus and stipe glabrous, odor indistinctive	_

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## **Additional information**

## **Conflict of interest**

The authors have declared that no competing interests exist.

## **Ethical statement**

No ethical statement was reported.

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# **Author contributions**

Data curation: YT. Investigation: SNL, YT. Resources: JBL, LGF. Validation: SNL, ARS, BMS. Writing - original draft: YXX. Writing - review and editing: ZXL, ZMT, LNL.

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# **Data availability**

All of the data that support the findings of this study are available in the main text.

# References

- Afyon A, Konuk M, Yağiz D, Helfer S (2005) A study of wood decaying macrofungi of the western Black Sea region, Turkey. Mycotaxon 93: 319–322.
- Aravindakshan DM, Manimohan P (2013a) A new species of *Mycena* sect. *Exornatae* and some taxonomic observations. Mycosphere 4(1): 146–150. https://doi.org/10.5943/mycosphere/4/1/9
- Aravindakshan DM, Manimohan P (2013b) *Mycena* sect. *Galactopoda*: Two new species, a key to the known species and a note on the circumscription of the section. Mycosphere 4(4): 653–659. https://doi.org/10.5943/mycosphere/4/4/1
- Aravindakshan DM, Kumar TKA, Manimohan P (2012) A new bioluminescent species of *Mycena* sect. *Exornatae* from Kerala State, India. Mycosphere 3(5): 556–561. https://doi.org/10.5943/mycosphere/3/5/4
- Aronsen A (1996) *Mycena juniperina*, a new member of section *Supinae* from Norway. Persoonia-Molecular Phylogeny and Evolution of Fungi 16(2): 257–259.
- Aronsen A, Læssøe T (2016) The genus *Mycena* s.l. Fungi of Northern Europe 5. Narayana Press, Gylling, Denmark, 373 pp.
- Baldrian P, Zrůstová P, Tláskal V, Davidová A, Merhautová V, Vrška T (2016) Fungi associated with decomposing deadwood in a natural beech-dominated forest. Fungal Ecology 23: 109–122. https://doi.org/10.1016/j.funeco.2016.07.001
- Ballesteros E (1984) Primera contribució al coneixement dels macromicets del Massís de Cadiretes (La Selva). Butlletí de la Institució Catalana d'Història Natural 51(Sec. Bot., 5): 67–76. https://digital.csic.es/bitstream/10261/97720/1/primera%20contribuci%C3%B3.pdf
- Bau T, Na Q, Liu LN (2021) A monograph of Mycenaceae (Agaricales) in China. Science press, Beijing, 326 pp.
- Beardslee HC, Coker WC (1924) The Mycenas of North Carolina. Journal of the Elisha Mitchell Scientific Society 40(1/2): 49–91. https://dc.lib.unc.edu/cdm/ref/collection/jncas/id/1348
- Bronckers R, Kelderman PH (2011) Paddenstoelen in Groeve't Rooth: een voorlopig resultaat van tien jaar inventariseren. Natuurhistorisch Maandblad 100(8): 121–128. https://natuurtijdschriften.nl/pub/1003301
- Casabón JCC (2015) El Género Lactarius En El Sur De Europa. Introducción Al Conocimiento De Algunas Especies Mediterráneas. Annali del XXI Giornate della Conferenza Europea di Micologia Mediterranea 21: 67–72. https://www.micobotanicajaen.com/AsoJaen/Actividades/Actividades2013/03CEMM2013/Anales.pdf
- Chen ZH, Zhang P (2019) Atlas of Macrofungi in Hunan. Hunan Science and Technology Press, Changsha, 426 pp.
- Chew ALC, Tan YS, Desjardin DE, Musa MY, Sabaratnam V (2014) Four new bioluminescent taxa of *Mycena* sect. *Calodontes* from Peninsular Malaysia. Mycologia 106(5): 976–988. https://doi.org/10.3852/13-274

- Chew ALC, Desjardin DE, Tan YS, Musa MY, Sabaratnam V (2015) Bioluminescent fungi from Peninsular Malaysia—a taxonomic and phylogenetic overview. Fungal Diversity 70(1): 149–187. https://doi.org/10.1007/s13225-014-0302-9
- Cho DH, Chung JY (2020) Study of fungal diversity of Mt. Cheonggye. Korean Journal of Nature Conservation 19(1): 29–60. https://doi.org/10.30960/kjnc.2020.19.1.29
- Cho DH, Chung JY (2023) Study of fungal diversity and fungal resources in Mt. Gwang-deok. Korean Journal of Nature Conservation 22(1): 1–26. https://doi.org/10.30960/kjnc.2023.22.1.1
- Cho M, Kwon SL, Jang S, Yoo Y, Lee SH, Kwon DY, Kim C, Lim YW, Kim JJ (2024) Notes of 15 unrecorded macrofungi in Korea. Journal of Species Research 13(1): 67–88. https://doi.org/10.12651/JSR.2024.13.1.067
- Cooper AC, Desjardin DE, Perry BA (2018) The genus *Mycena* (Basidiomycota, Agaricales, Mycenaceae) and allied genera from Republic of São Tomé and Príncipe, West Africa. Phytotaxa 383(1): 1–47. https://doi.org/10.11646/phytotaxa.383.1.1
- Corner EJH (1994) Agarics in Malesia. II Mycenoid. Beih. Nova Hedwig 109: 165-271.
- Cortés-Pérez A, Desjardin DE, Perry BA, Cruz V, Ramírez F, Villalobos-Arámbula AR, Rockefeller A (2019) New species and records of bioluminescent *Mycena* from Mexico. Mycologia 111(2): 1–20. https://doi.org/10.1080/00275514.2018.1554172
- Cortés-Pérez A, Guzmán-Dávalos L, Ramírez-Cruz V, Villalobos-Arámbula AR, Ruiz-Sanchez E, Ramírez-Guillén F (2023) New species of bioluminescent *Mycena* sect. *Calodontes* (Agaricales, Mycenaceae) from Mexico. Journal of Fungi 9(9): 902. https://doi.org/10.3390/jof9090902
- Desjardin DE (1995) A preliminary accounting of the worldwide members of *Mycena* sect. *Sacchariferae*. Bibliotheca Mycologica 159: 1–89.
- Desjardin DE, Perry BA, Lodge DJ, Stevani CV, Nagasawa E (2010) Luminescent *Mycena*: New and noteworthy species. Mycologia 102(2): 459–477. https://doi.org/10.3852/09-197
- Doğan HH, Karadelev M (2006) First record of *Mycena juniperina* from Turkey on a new host. Mycologia Balcanica 3: 77–79. https://doi.org/10.5281/zenodo.2547358
- Emmett EE (1992) British *Mycena* species—3. Mycologist 6(4): 165–173. https://doi.org/10.1016/S0269-915X(09)80559-4
- Fan LF, Wang BY, Ma TF, Li B, Ma JW, Lei XT, Bao NH (2024) Three new species of *Gymnopus* and *Mycena* (Agaricales, Basidiomycota) from Northwestern China. Frontiers in Microbiology 15: 1487598. https://doi.org/10.3389/fmicb.2024.1487598
- Fries EM (1821) Systema mycologicum, 3 Vols. Office Berlingiana, Lund, Greifswald, 202 pp.
- Fukasawa Y, Osono T, Takeda H (2009) Effects of attack of saprobic fungi on twig litter decomposition by endophytic fungi. Ecological Research 24(5): 1067–1073. https://doi.org/10.1007/s11284-009-0582-9
- Gao GM, Dou XY (1981) Hunan Physical Geography. Hunan People's Publishing House, Changsha, 204 pp.
- Gáperová S, Náplavová K, Gáper J (2015) Ectomycorrhizal and saprotrophic macrofungi associated with woody plants in the borova hora arboretum. Thaiszia Journal of Botany 25(1): 163–170.
- Gierczyk B, Kujawa A, Szczepkowski A, Ślusarczyk T, Kozak M, Mleczko P (2015) XXI Wystawa Grzybów Puszczy Białowieskiej: materiały do poznania mykobioty Puszczy Białowieskiej. Przegląd Przyrodniczy 26(3): 10–50.
- Gilliam MS (1976) The genus *Marasmius* in the northeastern United States and adjacent Canada. Mycotaxon 4(1): 1–144.

- Glez-Penñ D, Gómez-Blanco D, Reboiro-Jato M, Fdez-Riverola F, Posada D (2010) ALTER: Program-oriented conversion of DNA and protein alignments. Nucleic Acids Research 38(suppl\_2): W14–W18. https://doi.org/10.1093/nar/gkq321
- Grgurinovic CA (1998) *Mycena* in Australia: Section *Lactipedes*. Botanical Journal of Scotland 50(2): 199–208. https://doi.org/10.1080/03746609808684917
- Grgurinovic CA (2003) The Genus *Mycena* in South-Eastern Australia. Fungal Diversity Press, Hong Kong, 329 pp.
- Guerreiro MA, Kambach S, Stoll R, Brachmann A, Senker J, Begerow D, Peršoh D (2023) Linking processes to community functions—insights into litter decomposition combining fungal metatranscriptomics and environmental NMR profiling. Mycological Progress 22(2): 10. https://doi.org/10.1007/s11557-022-01859-0
- Gyosheva MM, Stoykov DY, Marinov YA (2016) Data on the fungal diversity of Balgarka Nature Park (Central Balkan, Bulgaria). Phytologia Balcanica 22(3): 309–322.
- Halama M, Romański M (2010) A new record of *Mycena picta* (Fr.: Fr.) Harmaja (Agaricales, Basidiomycota) from the Wigierski National Park (NE Poland). Opole Scientific Society Nature Journal 43: 29–36.
- Halama M, Chachuła P, Rutkowski R (2014) *Mycena Juniperina* (Agaricales, Basidiomycota), new for the polish and central European Mycobiota. Polish Botanical Journal 59(1): 109–116. https://doi.org/10.2478/pbj-2014-0008
- Hall TA (1999) BioEdit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nuclc Acids Symposium Series 41(41): 95–98. https://doi.org/10.12691/ajidm-4-3-3
- Harder CB, Lodge DJ, Petersen RH, Hughes KW, Blanco JC, Frøslev TG, Læssøe T (2012) Amyloidity is not diagnostic for species in the *Mycena pearsoniana* complex (*Mycena section Calodontes*). Mycological Progress 11: 725–732. https://doi.org/10.1007/s11557-011-0782-x
- Hintikka V (1963) Studies in the genus *Mycena* in Finland. Karstenia 6(7): 77–87. https://doi.org/10.29203/ka.1963.39
- Hughes KW, Petersen RH, Lickey EB (2009) Using heterozygosity to estimate a percentage DNA sequence similarity for environmental species' delimitation across basidiomycete fungi. New Phytologist 795–798. https://doi.org/10.1111/j.1469-8137.2009.02802.x
- Hughes KW, Matheny PB, Miller AN, Petersen RH, Iturriaga TM, Johnson KD, Methven AS, Raudabaugh DB, Swenie RA, Bruns TD (2020) Pyrophilous fungi detected after wildfires in the Great Smoky Mountains National Park expand known species ranges and biodiversity estimates. Mycologia 112(4): 677–698. https://doi.org/10.1080/00 275514.2020.1740381
- Imai S (1938) Studies on the Agaricaceae of Hokkaido. I. Journal of the Faculty of Agriculture. Hokkaido Imperial University 43(1): 1–178.
- Kalamees K, Raitviir A (2006) Macromycetes on Ainovy islands (Murmansk region, Russia). Meddelelser om Grønland. Bioscience 56: 139–147. https://doi.org/10.7146/mogbiosci.v56.142877
- Katoh K, Standley DM (2013) MAFFT multiple sequence alignment software version 7: Improvements in performance and usability. Molecular Biology and Evolution 30(4): 772–780. https://doi.org/10.1093/molbev/mst010
- Kirk PM, Cannon P, Stalpers J, Minter DW (2008) Dictionary of the Fungi. 10<sup>th</sup> edn. Centre for Agriculture and Bioscience International, Wallingford, UK, 771 pp.
- Kornerup A, Wanscher JH (1978) Methuen handbook of colour. Eyre Methuen, London, 252 pp.

- Kozlov AM, Darriba D, Flouri T, Morel B, Stamatakis A (2019) RAxML-NG: A fast, scalable, and user-friendly tool for maximum likelihood phylogenetic inference. Bioinformatics 35(21): 4453–4455. https://doi.org/10.1093/bioinformatics/btz305
- Kühner R (1926) Contribution à l'étude des Hyménomycètes et spécialement des Agaricacés. Le Botaniste 17: 1–215.
- Kühner R (1931) Utilisation de la réaction iodée dans la classification des *Mycena*. Bulletin Mensuel de la Societe Linneenne de Lyon 8: 122–127.
- Kühner R (1938) Le genre Mycena. Encyclopedie Mycologique 10: 1-710.
- Kwon SL, Jang S, Kim C, Lim YW, Kim JJ (2020) Note of five unrecorded mushrooms including three rare species on Mount Juwang in Korea. Mycobiology 48(3): 157–168. https://doi.org/10.1080/12298093.2020.1759348
- Laganà A, Salerni E, Barluzzi C, De Dominics V, Perini C (2002) Fungi (macromycetes) in various types of Mediterranean forest ecosystems (Tuscany, Italy). Polish Botanical Journal 47(2): 143–165.
- Lange JE (1914) Studies in the agarics of Denmark I. Dansk botanisk arkiv udgivet af Dansk botanisk forenin 1(5): 1–40.
- Lee HR, Han M, Choi MN, Lee H, Lee SW, Park EJ (2017) Enhancement of the germination efficiency of *Gastrodia elata* seeds using a new *Mycena* species. Journal of Plant Biotechnology 44(1): 56–60. https://doi.org/10.5010/JPB.2017.44.1.056
- Lee JK, Kim DH, Nguyen MH, Bae YJ, Manilak P (2021) A checklist of mushrooms of Dong Hua Sao National Biodiversity Conservation Area (DHSNBCA) of Lao-PDR. Journal of Forest and Environmental Science 37(2): 163–167. https://doi.org/10.7747/JFES.2021.37.2.163
- Li JZ, Hu XW, Peng YB (1993) Macrofungi of Hunan. Hunan Normal University Press, Changsha, 418 pp.
- Liu ZW, Na Q, Cheng XH, Wu XM, Ge YP (2021) *Mycena yuezhuoi* sp. nov. (Mycenaceae, Agaricales), a purple species from the peninsula areas of China. Phytotaxa 511(2): 148–162. https://doi.org/10.11646/phytotaxa.511.2.3
- Liu LN, Zhou GY, Tan ZM, Tian YX (2022a) Two new *Mycena* (Mycenaceae, Agaricales) species with rhizomorphs from subtropical areas of China. Phytotaxa 576(1): 75–88. https://doi.org/10.11646/phytotaxa.576.1.4
- Liu LN, Zhou GY, Shen AR, Shen BM, Tan Y, Tan ZM (2022b) *Mycena subpiligera* sp. nov., a symbiotic species from china associated with the seed germination of *Gastrodia elata*. Mycobiology 50(5): 294–301. https://doi.org/10.1080/12298093.20 22.2132001
- Liu XF, Deng PT, Wang XY, Zhang P (2024) A new edible *Clavulina* species from Hunan Province, China. Phytotaxa 661(2): 169–180. https://doi.org/10.11646/phytotaxa.661.2.4
- Liu LN, Shen AR, Shen BM, Tan Y, Li SN, Tan ZM (2025) Preliminary study on species diversity of symbiotic *Mycena* sensu lato promoting *gastrodia elata* seed germination. Acta Edulis Fungi 32(1): 65–76.
- Lu W, Priyashantha AKH, Galappaththi MC, Tibpromma S, Dai DQ, Patabendige NM, Premarathne BM, Kulasuriya DM, Ediriweera AN, Nimalrathna TS, Suwannarach N, Lumyong S, Tang A, Shao SC, Karunarathna SC (2024) Fungal bioluminescence: Past, present, and future. Diversity 16(9): 539. https://doi.org/10.3390/d16090539
- Łuszczyński J, Adamska E, Wojciechowska A, Czerwik-Marcinkowska J (2022) Diversity patterns of macrofungi in Xerothermic Grasslands from the Nida Basin (Małopolska Upland, Southern Poland): A case study. Biology 11(4): 531. https://doi.org/10.3390/biology11040531

- Maas Geesteranus RA (1980) Studies in Mycenas—15. A tentative subdivision of the genus *Mycena* in the northern Hemisphere. Persoonia-Molecular Phylogeny and Evolution of Fungi 11(1): 93–120. https://repository.naturalis.nl/pub/531705
- Maas Geesteranus RA (1983) Conspectus of the Mycenas of the Northern Hemisphere.
  - Sections sacchariferae, basipedes, bulbosae, clavulares, exiguae, and longisetae.
     Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen. Series
     Biological and Medical Sciences 86: 401–421.
- Maas Geesteranus RA (1986) Conspectus of the Mycenas of the Northern Hemisphere 6. Sections *Polyadelphia* and *Saetulipedes*. Koninklijke Nederlandse Akademie van Wetenschappen, Proceedings, Series C 89(2): 159–182.
- Maas Geesteranus RA (1988) Conspectus of the Mycenas of the Northern Hemisphere 9. Section *Fragilipedes*, species AG. Koninklijke Nederlandse Akademie van Wetenschappen, Proceedings, Series C 91: 43–83.
- Maas Geesteranus RA (1989) Conspectus of the Mycenas of the Northern Hemisphere, 12. Sections *Fuliginellae*, *Insignes*, *Ingratae*, *Euspeireae*, and *Caespitosae*. Koninklijke Nederlandse Akademie van Wetenschappen, Proceedings, Series C 92: 331–365.
- Maas Geesteranus RA (1992a) Mycenas of the Northern Hemisphere I. Studies in Mycenas and other papers. Koninklijke Nederlandse Akademie van Wetenschappen, Amsterdam, 493 pp.
- Maas Geesteranus RA (1992b) Mycenas of the Northern Hemisphere(II, Conspectus of the Mycenas of the Northern Hemisphere). Koninklijke Nederlandse Akademie van Wetenschappen, Amsterdam, 904 pp.
- Maas Geesteranus RA (1992c) Some Myceneae of the Himalayan foothills. Persoonia-Molecular Phylogeny and Evolution of Fungi 15(1): 33–53. https://archive.org/details/persoonia531739
- Maas Geesteranus RA, De Meijer AAR (1997) Mycenae paranaenses. Proceedings van de Koninklijke Nederlandse Akademie van Wetenschappen (Ser C), Amsterdam, North-Holland, 164 pp.
- Maas Geesteranus RA, Horak E (1995) *Mycena* and related genera from Papua new Guinea and new Caledonia. Bibliography. Mycology 159: 143–229.
- Malysheva V, Malysheva E, Voronina EY, Fedosova A, Bibikov N, Kiseleva D, Tiunov A, Kovalenko A (2017) Mycorrhiza of pyroloids (*Pyrola rotundifolia*, *P. media* and *Orthilia secunda*): Species composition of symbionts and trophic status of plants. Mikologiâ i Fitopatologiâ 51(6): 350–364.
- Marra D (2000) Osservazioni sui macromiceti nella Riserva Naturale Regionale del Lago di Villa (Challand St. Victor, Valle d'Aosta). Revue Valdôtaine d'Histoire Naturelle 54: 37–56.
- Matheny PB, Curtis JM, Hofstetter V, Aime MC, Moncalvo JM, Ge ZW, Yang ZL, Slot JC, Ammirati JF, Baroni TJ, Bougher NL, Hughes KW, Lodge DJ, Kerrigan RW, Seidl MT, Aanen DK, DeNitis M, Daniele GM, Desjardin DE, Kropp BR, Norvell LL, Parker A, Vellinga EC, Vilgalys R, Hibbett DS (2006) Major clades of Agaricales: A multilocus phylogenetic overview. Mycologia 98(6): 982–995. https://doi.org/10.1080/15572536.2006.11832627
- Métrod G (1949) Les Mycènes de Madagascar: (*Mycena, Corrugaria, Pterospora*). Prodrome à une flore mycologique de Madagascar 3: 1–146.
- Miyamoto T, Cha JY, Igarashi T (1996) A new record of *Mycena picta* from Japan. Mycoscience 37(4): 463–466. https://doi.org/10.1007/BF02461005
- Moncalvo JM, Vilgalys R, Redhead SA, Johnson JE, James TY, Aime MC, Valerie H, Verduin SJW, Larsson E, Baroni TJ, Thorn RG, Jacobsson S, Clémençon H, Miller OK (2002) One hundred and seventeen clades of euagarics. Molecular Phylogenetics and Evolution 23(3): 357–400. https://doi.org/10.1016/S1055-7903(02)00027-1

- Moreno G, Albertó E (1996) Agaricales sensu lato de Argentina. I. Cryptogamie Mycologie 17(2): 61–84. https://doi.org/10.5962/p.354462
- Mustafabayli EH, Aghayeva DN (2019) Mushroom diversity in Shaki district of Azerbaijan. Plant & Fungal Research 2(1): 23–31. https://doi.org/10.29228/plantfungalres.45
- Na Q, Bau T (2018) New species of *Mycena* (Mycenaceae, Agaricales) with colored lamellae and three new species records from China. Phytotaxa 361(3): 266–278. https://doi.org/10.11646/phytotaxa.361.3.2
- Na Q, Bau T (2019a) *Mycena* section *Sacchariferae*: Three new species with basal discs from China. Mycological Progress 18: 483–493. https://doi.org/10.1007/s11557-018-1456-8
- Na Q, Bau T (2019b) Recognition of *Mycena* sect. *Amparoina* sp. nov. (Mycenaceae, Agaricales), including four new species and revision of the limits of sect. *Sacchariferae*. MycoKeys 52: 103–124. https://doi.org/10.3897/mycokeys.52.34647
- Na Q, Liu ZW, Zeng H, Ke BR, Song ZZ, Cheng XH, Ge YP (2022) Taxonomic studies of bluish *Mycena* (Mycenaceae, Agaricales) with two new species from northern China. MycoKeys 90: 119. https://doi.org/10.3897/mycokeys.90.78880
- Nauta M, Vellinga EC (1992) Towards a distribution atlas of macrofungi in the Netherlands. Mycologist 6(1): 6–10. https://doi.org/10.1016/S0269-915X(09)80506-5
- Nylander J (2004) MrModeltest 2.3, Computer program and documentation distributed by the author. Evolutionary Biology Centre, Uppsala University, Uppsala.
- Oliveira JJ, Vargas-Isla R, Cabral TS, Cardoso JS, Andriolli FS, Rodrigues DP, Ikeda T, Clement CR, Ishikawa NK (2021) The Amazonian luminescent *Mycena cristinae* sp. nov. from Brazil. Mycoscience 62(6): 395–405. https://doi.org/10.47371/mycosci.2021.05.004
- Osmundson TW, Robert VA, Schoch CL, Baker LJ, Smith A, Robich G, Mizzan L, Garbelotto MM (2013) Filling gaps in biodiversity knowledge for macrofungi: Contributions and assessment of an herbarium collection DNA barcode sequencing project. PLoS ONE 8(4): e62419. https://doi.org/10.1371/journal.pone.0062419
- Pegler DN (1977) A preliminary agaric flora of East Africa. Kew Bulletin Additional Series 6: 1–615.
- Pegler DN (1983) Agaric flora of the Lesser Antilles. Kew Bulletin Additional Series 9: 1–668.
- Pegler DN (1986) Agaric Flora of Sri Lanka. Kew Bulletin Additional Series 12: 1-519.
- Pegler DN (1987) A revision of the Agaricales of Cuba 1. Species described by Berkeley & Curtis. Kew Bulletin 42(3): 501–585. https://doi.org/10.2307/4110064
- Perry BA (2002) A taxonomic investigation of *Mycena* in California. Doctoral dissertation, San Francisco State University, California, USA
- Perry BA, Desjardin DE (2016) New species of *Mycena* (Basidiomycota, Agaricales) from California. Phytotaxa 269(1): 33–40. https://doi.org/10.11646/phytotaxa.269.1.4
- Petersen RH, Hughes KW, Lickey EB, Kovalenko AE, Morozova OV, Psurtseva NV, Morosova O (2008) A new genus, *Cruentomycena*, with *Mycena viscidocruenta* as the type species. Mycotaxon 105(4): 119–136.
- Qing CJ (1990) Hunan vegetation. Hunan Science and Technology Press, Changsha, 420 pp.
- Rathnayaka AR, Tennakoon DS, Jones GE, Wanasinghe DN, Bhat DJ, Priyashantha AH, Stephenson SL, Tibpromma S, Karunarathna SC (2024) Significance of precise documentation of hosts and geospatial data of fungal collections, with an emphasis on plant-associated fungi. New Zealand Journal of Botany 63(2–3): 462–489 [1–28]. https://doi.org/10.1080/0028825X.2024.2381734

- Retnowati A, Kusumawaty A, Apandi I, Haryadi A (2020) Two newly recorded species of Agaricales (Basidiomycota) from Banggai Kepulauan, Sulawesi, Indonesia. Biodiversitas Journal of Biological Diversity 21(12): 5615–5621. https://doi.org/10.13057/biodiv/d211217
- Robich G (2003) *Mycena* d'Europa Vol. 1. Associazione Micologica Bresadola, Trento, Italy, 728 pp.
- Robich G (2016) *Mycena* d'Europa Vol. 2. Associazione Micologica Bresadola, Trento, 796 pp.
- Ronquist F, Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19(12): 1572–1574. https://doi.org/10.1093/bioinformatics/btg180
- Roussel HFA (1806) Flore du Calvades et des terreins adjacens. 2<sup>nd</sup> edn. Caen, 252 pp.
- Rudolf K, Pál-Fám F, Morschhauser T (2016) Macrofungi records from a wood pasture in the Belső-Cserehát (NE Hungary). Studia Botanica Hungarica 47(1): 29–40. https://doi.org/10.17110/StudBot.2016.47.1.29
- Senn-Irlet B (1987) Pilze aus der alpinen stufe des Val d'Anniviers (Wallis). Bulletin de la Murithienne 105: 87–106. https://core.ac.uk/reader/20655250
- Senn-Irlet B, Mürner R, Martini E, Küffer N, Marchi RD, Bieri G (2012) Saprobic fungi on wood and litter of Alnus alnobetula in the Swiss Alps. Mycotaxon 120(506): 1–34.
- Shiryaeva OS (2018) New records of agaricoid fungi from Sverdlovsk Region, Russia. Botanica Lithuanica 24(2): 150–161. https://doi.org/10.2478/botlit-2018-0015
- Singer R (1986) The Agaricales in modern taxonomy. Koeltz Scientific Books, Koenigstein, 1609 pp.
- Singer R (1989) New taxa and new combinations of Agaricales (diagnoses fungorum novorum Agaricalium IV). Fieldiana: Botany New Series 21: 1–133. https://doi.org/10.5962/bhl.title.2537
- Singer R, Digilio AP (1951) Pródromo de la flora agaricina Argentina. Lilloa 25: 1-461.
- Smith AH (1935) Studies in the genus *Mycena*. I. American Journal of Botany 22(10): 858–877. https://doi.org/10.1002/j.1537-2197.1935.tb05071.x
- Smith AH (1947) North American Species of *Mycena*. University of Michigan Press, Ann Arbor, Michigan, 521 pp.
- Swartz D (1933) Studies of Arkansas Fungi. I. Basidiomycetes. American Midland Naturalist 14(6): 714–719. https://doi.org/10.2307/2420128
- Swindell SR, Plasterer TN (1997) Sequence data analysis guidebook. Humana Press, Totowa.
- Tholl MT, Baral HO, Schultheis B, Marson G, Diederich P (2000) Journées luxembourgeoises de mycologie vernale 1998. Bulletin de la Société des Naturalistes Luxembourgeois 100: 39–62. http://lichenology.info/pdf/Tholl2000.pdf
- Türkekul İ (2017) New *Calbovista, Mycena, Rhizopogon, Stictis*, and *Symphyosirinia* records from Turkey. Mycotaxon 132(3): 503–512. https://doi.org/10.5248/132.503
- Vaidya G, Lohman DJ, Meier R (2011) SequenceMatrix: Concatenation software for the fast assembly of multi-gene datasets with character set and codon information. Cladistics 27(2): 171–180. https://doi.org/10.1111/j.1096-0031.2010.00329.x
- Veerkamp MT (2001) Paddestoelen in acht bosreservaten; Stille Eenzaamheid, Kremboong, Tongerense Hei, Norgerholt, Zwarte Bulten, Mattemburgh, Hollandse Hout en Houtribbos. Alterra, Wageningen, 101 pp.
- Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. Journal of Bacteriology 172(8): 4238–4246. https://doi.org/10.1128/jb.172.8.4238-4246.1990

- Villarreal M, Couceiro A, Sos C, Mateos J, Rodrigues H (2024) *Mycena dasypus* (Agaricales, Mycenaceae), Primer Registro En La Península Ibérica. Boletin de la Sociedad Micologica de Madrid 48: 63–70.
- Vishwakarma P, Tripathi NN, Singh P (2017) A checklist of macrofungi of Gorakhpur District, UP India. Current Research in Environmental & Applied Mycology 7(2): 109–120. https://doi.org/10.5943/cream/7/2/8
- Vu D, Groenewald M, De Vries M, Gehrmann T, Stielow B, Eberhardt U, Al-Hatmi A, Groenewald JZ, Cardinali G, Houbraken J, Boekhout T, Crous PW, Robert V, Verkley GJM (2019) Large-scale generation and analysis of filamentous fungal DNA barcodes boosts coverage for kingdom fungi and reveals thresholds for fungal species and higher taxon delimitation. Studies in Mycology 92(1): 135–154. https://doi.org/10.1016/j.simyco.2018.05.001
- Watling R (1984) Larger fungi around Kindrogan, Perthshire. Transactions of the Botanical Society of Edinburgh 44(3): 237–259. https://doi.org/10.1080/03746608408685391
- Wei RX, Ge YP, Qi LL, Han MH, Zeng H, Hu YP, Zou L, Cheng XH, Wu XM, Na Q (2024) Revealing brownish *Mycena* diversity in China: new discoveries and taxonomic insights. Journal of Fungi 10(6): 439. https://doi.org/10.3390/jof10060439
- White T, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis M, Gelfand D, Shinsky J, White T (Eds) PCR protocols: a guide to methods and applications, 315–322. https://doi.org/10.1016/B978-0-12-372180-8.50042-1
- Zhang P, Deng HZ, Chen ZH, Li JZ (2005) Atlas of Macrofungi in Hupingshan, Hunan. Hunan Science and Technology Press, Changsha, 325 pp.
- Zhang H, Xiao YX, Tan ZM, Shen AR, Shen BM, Tan Y, Li SN, Feng LG, Liu ZX, Liu LN (2024) *Mycena brunnescens* (Basidiomycota, Mycenaceae), a new species of *Mycena* sect. *Pterigenae* from China. Biodiversity Data Journal 12: e125570. https://doi.org/10.3897/BDJ.12.e125570